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Three-dimensional printing of Hela cells for cervical tumor model *in vitro*

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Abstract

Advances in three-dimensional (3D) printing have enabled the direct assembly of cells and extracellular matrix materials to form *in vitro* cellular models for 3D biology, the study of disease pathogenesis and new drug discovery. In this study, we report a method of 3D printing for Hela cells and gelatin/alginate/fibrinogen hydrogels to construct *in vitro* cervical tumor models. Cell proliferation, matrix metalloproteinase (MMP) protein expression and chemoresistance were measured in the printed 3D cervical tumor models and compared with conventional 2D planar culture models. Over 90% cell viability was observed using the defined printing process. Comparisons of 3D and 2D results revealed that Hela cells showed a higher proliferation rate in the printed 3D environment and tended to form cellular spheroids, but formed monolayer cell sheets in 2D culture. Hela cells in 3D printed models also showed higher MMP protein expression and higher chemoresistance than those in 2D culture. These new biological characteristics from the printed 3D tumor models *in vitro* as well as the novel 3D cell printing technology may help the evolution of 3D cancer study.

Keywords: 3D cell printing, bioprinting, tumor models, *in vitro* tumor model, cancer model

(Some figures may appear in colour only in the online journal)

1. Introduction

Cancer is one of the most serious diseases, with over 10 million new cases diagnosed worldwide each year. Despite many efforts, an inadequate understanding of tumorigenesis still hinders the development of cancer therapy [1]. Although the most effective way of studying tumors and testing anti-tumor drugs is in clinical trials, ethical and safety limitations prevent this method from being widely used. To overcome this hurdle, preclinical tumor models are often used to mimic physiological environments of tumors for tumorgenesis study and anti-cancer drug screening [2–4]. For example, Jordan *et al* used two-dimensional (2D) monolayered Hela cell cultures to study the chemotherapy mechanism of the anti-tumor drug paclitaxel (also known as Taxol) [3]. Ellingsen *et al* used surgical specimen xenograft models (animal models) to mimic the physiological microenvironment of cervical carcinoma and studied the mechanism of tumor metastasis and radiation sensitivity [4]. However, 2D monolayered cellular models

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lack the microenvironment characteristics of natural three-dimensional (3D) tissues *in vivo* [5]. On the other hand, animal models established in immunocompromised mice may show false effects on tumor development and progression [1]. To overcome these hurdles, *in vitro* 3D tumor models based on human cancer cells have been increasingly used in order to accurately reproduce the characteristics of human cancer tissues [6, 7]. Relevant studies of biological characteristics for cell proliferation [8], morphology [9], drug metabolism [10], gene expression and protein synthesis [11] were reported for 3D tumor models which compared with 2D planar culture. Various techniques, such as multicellular spheroids [12–14], cell-seeding 3D scaffolds [11, 15], hydrogel embedding [16, 17], microfluidic chips [18, 19] and cell patterning [20, 21] have also been developed for construction of 3D *in vitro* tumor models. For example, Ridky *et al* reported that the gene alterations in spontaneous tumors were similar to the 3D organotypic tissues in the constructed 3D organotypic tissues seeded with epithelial cells onto basement membranes comparing with the 2D planar cell culture [11]. Loessner *et al* discovered that ovarian cancer cell lines showed higher chemoresistance in 3D hydrogels than in 2D culture [16]. Liu *et al* engineered a microfluidic 3D co-culture tumor model of cancer cells and carcinoma-associated fibroblasts and found increased cancer cell invasion in the 3D microfluidic channels with fibroblast co-culture [19]. Xu *et al* patterned ovarian carcinoma cells and human diploid fibroblast cells onto a 2D Matrigel surface and demonstrated the formation of 3D cellular acini [21]. Although these studies revealed useful characteristics of 3D tumor models *in vitro*, to our knowledge, it is still difficult to simulate a complex 3D physiological tumor microenvironment in most of the above models due to the limitations of the fabrication techniques.

Advances in 3D printing have enabled direct assembly of cells and extracellular matrix (ECM) materials to form *in vitro* cellular models for 3D biology, the study of disease pathogenesis and new drug discovery. This promising technique has offered an opportunity for the biofabrication of complex 3D *in vitro* models with simulated physiological microenvironments [22]. The application of 3D cell printing has been reported in the printing of 3D large-scale tissue constructs [23], *in vitro* liver tissues [24], adipose tissues [25], bone tissues [26] and hybrid tissue constructs with vascular-like networks [27]. The objective of this paper is to report our study on the construction and characterization of *in vitro* cervical tumor models by 3D printing of Hela cells (cervical tumor cells) and gelatin/alginate/fibrinogen hydrogel biomaterials. Since the Hela cell line derived from cervical cancer cells was among the first cell lines successfully cultured *in vitro* and was widely used for tumor studies [28], we chose to use Hela cells for this study. In addition, since native ECM consists of fibrous proteins such as fibronectin, collagen and laminin [29], hydrogels such as alginate [30], fibrin [31] and gelatin [32] are widely used as ECM mimics for cell/tumor cell culture. For this reason we used these materials in this study to print 3D tumor constructs in order to mimic the ECM characteristics and cervical cancer microenvironment. The printing technique and the method of construction of the 3D tumor models will be introduced. The results of biological characterization of cell proliferation, matrix metalloproteinase (MMP) protein expression and chemoresistance for the printed 3D cervical tumor models will be presented and also compared with conventional 2D planar culture models.

2. Materials and methods

2.1. Cell culture

The Hela cells were obtained from the Center for Animal Experiments/A3 Lab in Wuhan University. The cells were cultured in high-glucose Dulbecco's modified Eagle medium (H-DMEM; Invitrogen) with 10% fetal bovine serum (FBS; Hyclone) in a CO_2 incubator at 37 °C and with 5% CO_2. The Hela cells were subcultured by trypsin (0.25%; invitrogen) dissociation at about 80% confluence. The culture media were changed every 2–3 days.

2.2. Material preparation

Gelatin powder (Sigma; G1890) was dissolved in 0.9% NaCl solution (w/v) at 20% (w/v). Sodium alginate powder (Sigma; A0682) was dissolved in 0.9% NaCl solution (w/v) at 4% (w/v). Both solutions were sterilized by heating three times in a stove (70 °C) for 30 min. Fibrinogen (Sigma; F8630) was dissolved in H-DMEM at 8% (w/v).

2.3. Construct fabrication and culture

A 3D cell printer developed by our group (cell assembly system I) was used to fabricate 3D tumor-like constructs similar to those previously described [23, 27]. Briefly, Hela cells were collected by centrifuge at 1000 r min^{-1} for 5 min and suspended in an 8% fibrinogen solution to a density of 4×10^6 cells mL^{-1}. A fibrinogen/Hela mixture, 20% gelatin solution and 4% sodium alginate solution were evenly mixed at a volume ratio of 1:2:1. Finally, the mixture was composed of 10% gelatin, 1% sodium alginate, 2% fibrinogen and Hela cells with a density of 10^6 cells mL^{-1}. One milliliter of the cell/biomaterial mixture was drawn into a sterilized commercial syringe with a 25 gauge needle. The mixture was physically crosslinked at 25 °C for about 5 min in the syringe and then mounted onto the 3D cell printer. A Hela/hydrogel construct with a grid structure of $10 \times 10 \times 2$ mm^3 was fabricated by forced extrusion in a sterile atmosphere of 10 °C in a layer-by-layer fashion. $CaCl_2$ (3%, w/v) was gently added to chemically crosslinked alginate in the 3D constructs. The construct was then immersed in 20 U mL^{-1} thrombin (Sigma; T4648) for 15 min to crosslink fibrinogen. Between each solution addition, the constructs were gently washed in phosphate buffered saline (PBS) two to three times (figure 1(*A*)). Each construct was cultured in a 35 mm petri dish with 2 mL culture media per dish.

A 2D planar culture sample was prepared by seeding Hela at a density of 5000 cells cm^{-2} in 35 mm petri dishes with 2 mL culture media per dish. Both 3D Hela/hydrogel constructs and 2D samples were cultured in H-DMEM supplemented with 20 mg L^{-1} aprotinin (YEASEN) and 10% FBS at 37 °C

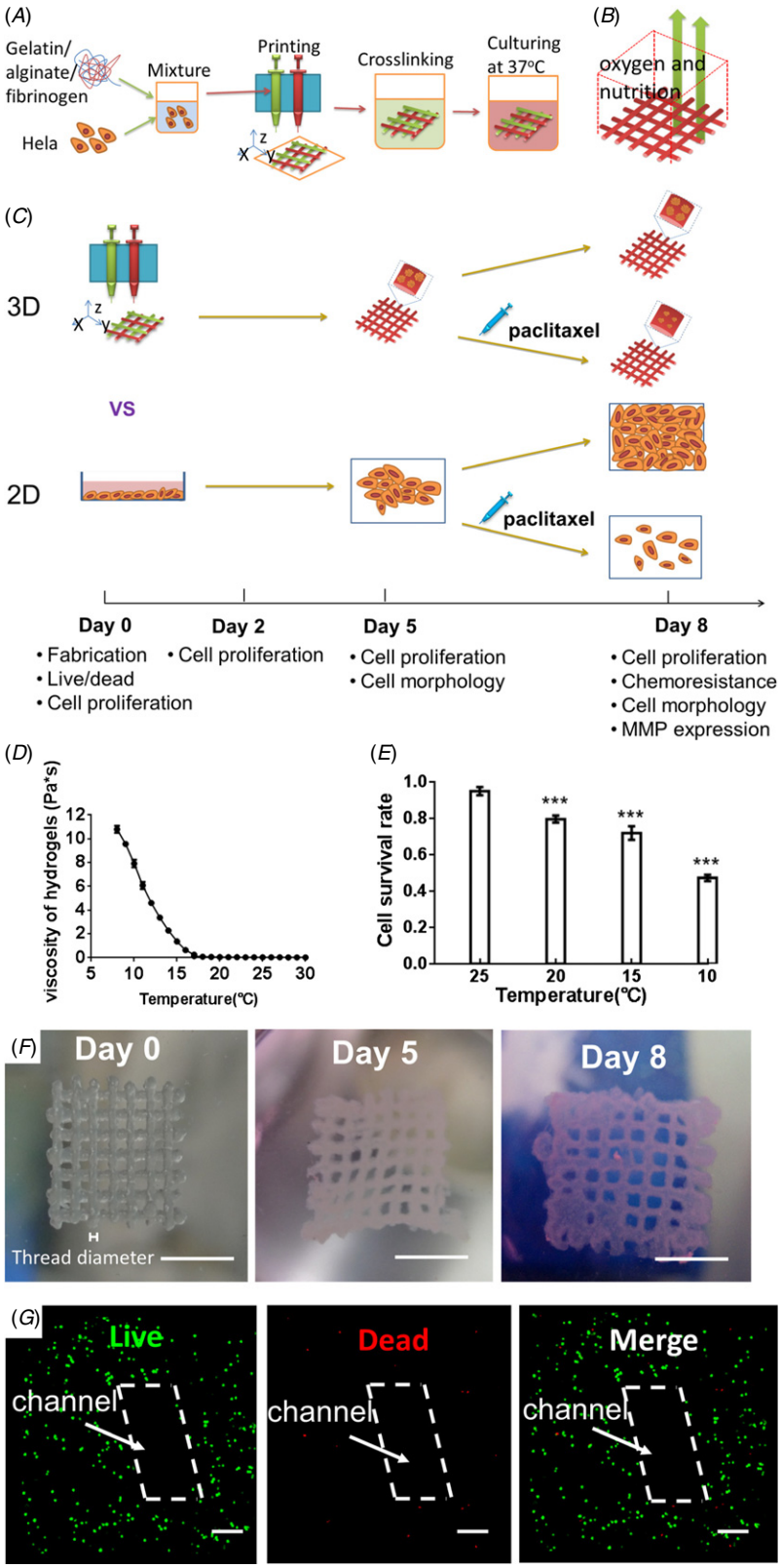

Figure 1. Fabrication of 3D Hela/hydrogel constructs. (*A*) Schematic of the 3D cell printing process. (*B*) The design of the 3D Hela/hydrogel constructs. (*C*) Schematic of the timeline of this research. Both 3D Hela/hydrogel constructs and 2D planar samples were cultured for 5 days and 3 more days with/without paclitaxel addition. (*D*) The viscosity of hydrogels at different temperatures. (*E*) Cell survival rate at different temperatures. *** means $p < 0.001$; *t*-test. (*F*) Top view of 3D Hela/hydrogel constructs on day 0, day 5 and day 8. Scale bar, 5 mm. (*G*) Cell viability after printing by live/dead staining under LSCM, where live cells are stained in green and dead cells are stained in red. Scale bar, 200 μm.

with 5% CO_2 for 8 days. Aprotinin, a proteinase inhibitor, was dissolved in the culture medium to inhibit the fibrin degradation and keep the constructs stable [33]. The culture media were changed on day 2 and 5. The culture medium was collected and centrifuged at 1000 r min^{-1} for 5 min for MMP expression tests on day 8. The timeline of this research is shown in figure 1(*B*).

2.4. Viscosity of Hela/hydrogels

Before crosslinking (as described in section 2.3) the Hela/hydrogel mixture was added into the rotational rheometer (MCR302, Anton Paar) to analyze the viscosity. The shear rate was constant (100 s^{-1}) and the temperature changed from 30 °C to 8 °C. Then the viscosity was recorded every 5 s. Three independent samples were tested.

2.5. Cell survival rate

Three independent samples were printed at different nozzle temperatures. Cell survival rate in the 3D Hela/hydrogel constructs was assessed immediately after biofabrication to determine the influence of the printing process on cell viability. A fluorescent live/dead staining was carried out according to the manufacturer's instructions. Briefly, the mixture of Calcein-AM (Dojindo; 1 μmol mL^{-1}) and PI (Sigma; 2 μmol mL^{-1}) was filtered through a 0.22 mm filter prior to staining. Hela/hydrogel constructs were stained by incubation with a Calcein AM-PI mixture for 30 min in the dark at 37 °C and gently washed three times with PBS. A laser scanning confocal microscope (LSCM; LSM710META, ZEISS) was used for image acquisition. Cell viability was calculated as (number of green stained cells/number of total cells) × 100%. Three random fields were chosen for each sample. Three independent samples were counted.

2.6. Cell proliferation analysis

The cell counting kit-8 (CCK-8; Dojindo) was used to analyze cell proliferation of both 3D Hela/hydrogel constructs and 2D planar culture samples on days 0, 2, 5 and 8 according to the manufacturer's instructions. Briefly, both 3D Hela/hydrogel constructs and 2D planar culture samples were washed with PBS three times. Then 1 mL H-DMEM and 0.1 mL CCK-8 solution was added into each 35 mm petri dish. After 2 h of incubation at 37 °C, 0.5 mL culture medium was transferred to a 96-well plate and read by fluorescence with an excitation 450 nm and emission 630 nm filter pair (Model680, Bio-Rad). 3D Hela/hydrogel constructs without cells and petri dishes without cells were subjected to the same process to use as blanks. The data of both 3D Hela/hydrogel constructs and 2D planar culture samples were normalized to day 0 (4 h after biofabrication or 2D cell seeding). Three independent samples were tested in each group.

2.7. Cell morphology imaging and analysis

A phase-contrast microscope (DP70, Olympus) was used to observe and record cell morphology during the whole experimental process. Staining of f-actin filaments and cellular nuclei was also applied to determine cellular morphological change in 3D Hela/hydrogel constructs and 2D planar culture. Briefly, samples were washed with PBS three times, fixed with 4% paraformaldehyde for 20 min, permeabilized for 30 min by 0.1% Triton X-100, blocked with 1% bovine serum albumin for 30 min and then stained with FITC-phalloidin (5 μg mL^{-1}; Sigma) for 20 min at room temperature with light avoidance. Cell nuclei were stained with DAPI (1 μg mL^{-1}; Sigma) for 5 min at room temperature with light avoidance. Samples were washed with PBS three times between incubations. LSCM (LSM710META, ZEISS) was used for image acquisition.

To quantify morphological changes of Hela in the 3D model, phase-contrast microscope images were quantitatively analyzed by Image Pro Plus software. Three images at three different positions were measured. LSCM images were quantitatively analyzed using ZEN 2009 software. Four different positions with more than ten cellular spheroids in each position were measured.

2.8. Protein characterization

The collected supernatant for MMP expression described in section 2.3 was mixed with loading buffer and loaded onto 12% acrylamide gel containing gelatin. Protein concentrations were determined using a BCA kit (Pierce, Rockford, IL) for normalizing the protein amount of 3D Hela/hydrogel constructs and 2D planar culture samples. Gel was run at 110 V for 100 min (Electrophoresis System, Bio-Rad) and then treated by an MMP Zymography Assay Kit (Applygen) according to the standard protocol. Gel was then stained in Coomassie brilliant blue staining buffer for 3 h, de-stained until clear bands were visible and semi-quantified by Quantity One software. Three independent samples were tested in each group.

2.9. Chemoresistance test

As shown in figure 1(*B*), three independent 2D and 3D samples were randomly picked out on day 5 for chemoresistance studies. Briefly, samples were cultured in H-DMEM supplemented with 20 mg L^{-1} aprotinin, 10% FBS and 50 μg L^{-1} paclitaxel (Gene Operation) for another 3 days, and tested using a CCK-8 kit as described in section 2.5.

2.10. Statistical analysis

Statistical analysis was performed by GraphPad Prism using two-way analysis of variance (ANOVA) in conjugation with a Bonferroni post-hoc test and a Student t-test. Differences were considered statistically significant when p values were lower than 0.05. All data are presented as mean ± standard deviation. Three independent trials were carried out unless otherwise stated.

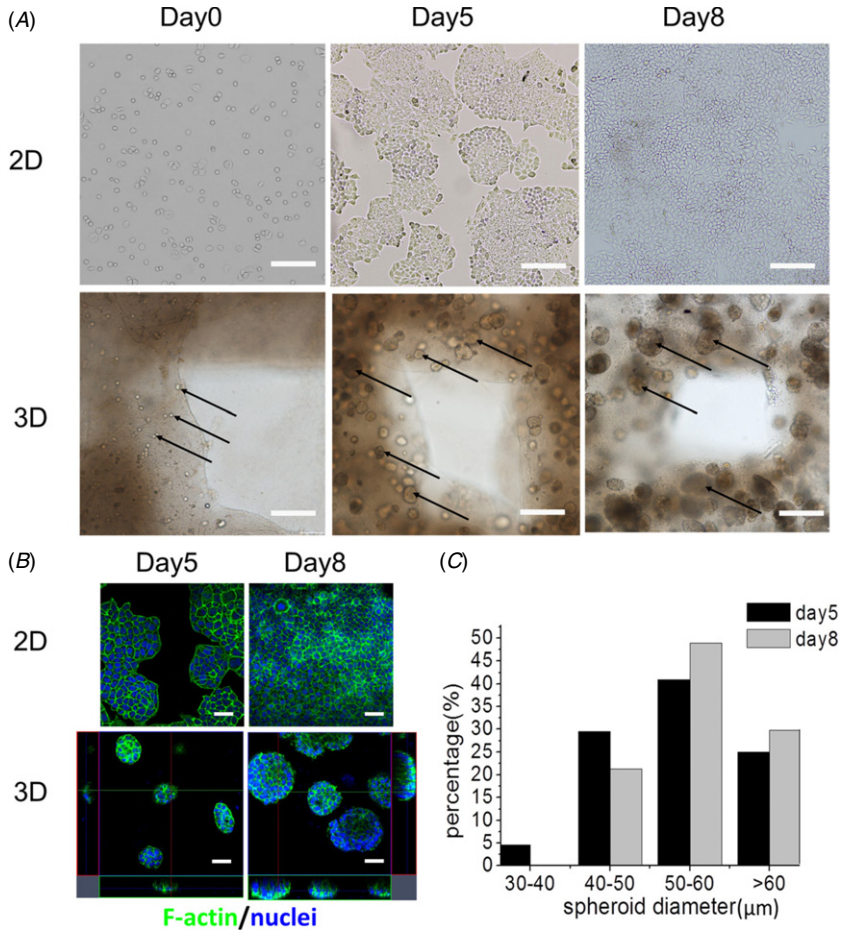

Figure 2. Cellular morphological changes during 8 days of culture in 3D constructs and 2D planar culture. (*A*) Hela cells in 2D planar culture and 3D Hela/hydrogel constructs observed by a phase-contrast microscope on day 0, day 5 and day 8. Scale bar, 200 μm. Black arrows indicate cells and cellular spheroids in the 3D construct. (*B*) Cytoskeleton distribution by staining on day 5 and day 8 in 2D planar culture and 3D constructs observed under LSCM. Scale bar, 50 μm. (*C*) Distribution of spheroid diameter in 3D Hela/hydrogel constructs on day 5 and day 8.

3. Results

3.1. Fabrication of 3D tumor-like constructs

In order to construct 3D tumor models *in vitro* to mimic natural cervical tumors, we used cell printing technology to fabricate 3D Hela/gelatin/alginate/fibrinogen constructs. A cuboid structure with interconnected channels (figure 1(*C*)) was designed to allow the transport of nutrients, oxygen and metabolic waste. As shown in figure 1(*D*), the viscosity of Hela/hydrogel increased with the decline of hydrogel temperature (30 °C to 10 °C), and the viscosity of Hela/hydrogel increased significantly at the temperature range from 20 °C to 10 °C. The cell survival rate decreased with the decline of the nozzle temperature from 25 °C to 10 °C, with significant differences (figure 1(*E*)). We chose the parameters of 10 mm^3 min^{-1} extrusion speed, 250 μm nozzle inner diameter, 10 °C chamber temperature and 25 °C nozzle temperature to print 3D Hela/hydrogel constructs. The printed 3D Hela/hydrogel constructs showed a clear and stable structure with interconnected channels. The fibers of 3D Hela/hydrogel constructs were uniform and smooth with a mean thread diameter of 500 μm (figure 1(*F*)). Hela/hydrogel constructs maintained good structural stability for 8 days (figure 1(*F*)). After printing, the viability of Hela cells in the constructs was 94.9% $\pm$ 2.2% (figure 1(*G*)).

3.2. Cellular morphological change in 3D constructs and 2D culture

A phase-contrast microscope was used to observe the morphology of Hela cells over an 8 day experimental period in the tumor-like constructs, as shown in figure 2(*A*). Compared with 2D planar culture, Hela cells in 3D Hela/hydrogel constructs showed a spheroid morphology on day 5, and their diameters continued to grow until day 8. Based on semi-quantitative analysis of phase-contrast microscope images, 79.5% $\pm$ 6.8% areas of hydrogel were taken up by Hela spheroids. To further analyze cellular morphology in the 3D constructs, cell filaments and nuclei were visualized by staining and observed under LSCM, as shown in figure 2(*B*). It was demonstrated that Hela cells formed round spheroids with smooth surfaces and tight cell-cell connections within the 3D hydrogel, whereas Hela cells cultured on 2D tissue culture plates showed a flat and elongated morphology. Image-based semi-quantitative analysis showed increased spheroid

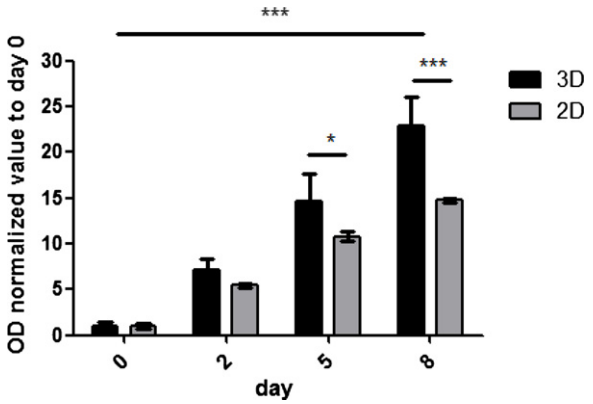

Figure 3. Cellular metabolic change in 2D planar culture and 3D constructs from day 0 to day 8. * means $p < 0.05$, *** means $p < 0.001$; ANOVA in conjugation with Bonferroni *post-hoc* test.

diameter from day 5 (44 μm $\pm$7 μm) to 8 (58 μm $\pm$ 10 μm) (figure 2(*C*)).

3.3. Cell proliferation and MMP expression

To determine cell proliferation in 3D tumor-like constructs and 2D culture, a CCK-8 kit was used to analyze cellular metabolic activity on day 0, 2, 5 and 8. 2D samples were treated under the same protocol as 3D constructs. It was demonstrated that, compared with day 0, Hela cells in the 2D planar culture showed 5.4-fold proliferation on day 2, 10.8-fold proliferation on day 5 and 14.8-fold proliferation on day 8, while Hela cells in 3D Hela/hydrogel constructs showed 7.2-fold proliferation on day 2, 14.6-fold proliferation on day 5 and 22.8-fold proliferation on day 8. There were significant differences between 3D and 2D samples on day 5 and 8 (figure 3).

MMP-2 and MMP-9 secretion was analyzed using an MMP Zymography assay kit to determine whether the 3D hydrogel environment affected Hela MMP secretion. As shown in figure 4(*A*), the bands of both MMP-2 and MMP-9 in 3D Hela/hydrogel constructs were brighter compared with those in the 2D planar culture. Semi-quantitative grayscale analysis of the bands further indicated that MMP-9 and MMP-2 secretion in 3D construct was 2.3 times and 2.5 times that of the 2D sample, with significant differences (figure 4(*B*)).

3.4. Chemoresistance

Paclitaxel was added into the culture media of the 3D tumor-like constructs and 2D culture samples and incubated for three days to analyze chemoresistance of Hela cells in different conditions. Abundant cellular apoptosis was observed in both the 2D cell culture and 3D constructs after the addition of paclitaxel. The cell morphology became irregular and the cytoskeleton showed loosened morphology (figure 5(*A*)). Most cells in the 2D culture floated from the substrate, while cellular spheroids in the 3D Hela/hydrogel constructs were still maintained within the hydrogel threads.

Figure 5(*B*) demonstrates a dramatic decline of cellular metabolic activity after the addition of paclitaxel. Compared with day 5, which was when paclitaxel was first added, the metabolic activity declined to 0.74 and 0.09 times in the 3D and 2D samples, respectively, with significant differences. Compared with the positive control, which was cultured in H-DMEM without adding paclitaxel, the paclitaxel-added samples showed 0.47 and 0.06 times the metabolic activity in 3D and 2D culture, respectively, with significant differences.

Semi-quantitative analysis of cellular spheroid diameters (figure 5(*C*)) showed a largely declined mean diameter and non-uniformed distribution in the paclitaxel group (40 μm $\pm$ 14 μm) compared with the non-paclitaxel group (58 μm $\pm$ 10 μm).

4. Discussion

3D tumor models with microenvironmental characteristics of cell–cell and cell–matrix interactions *in vivo* are becoming important tools for drug testing and tumor biological studies [1, 2, 5]. In this study, we printed 3D Hela/hydrogel constructs as cervical tumor models, and 3D tumor characteristics were studied.

Cell survival rate is one of the key factors to consider while applying 3D cell printing technology in the construction of tissue-like models. Cells are subjected to mechanical forces during the 3D extrusion cell printing process. It is well known that increased mechanical forces cause cellular damage and thus reduce cell survival rate [34]. Mechanical forces in the 3D cell printing process are determined by parameters like extrusion speed, nozzle diameter, viscosity of hydrogels, chamber temperature and nozzle temperature. Decreasing the nozzle diameter, increasing the extrusion speed and increasing

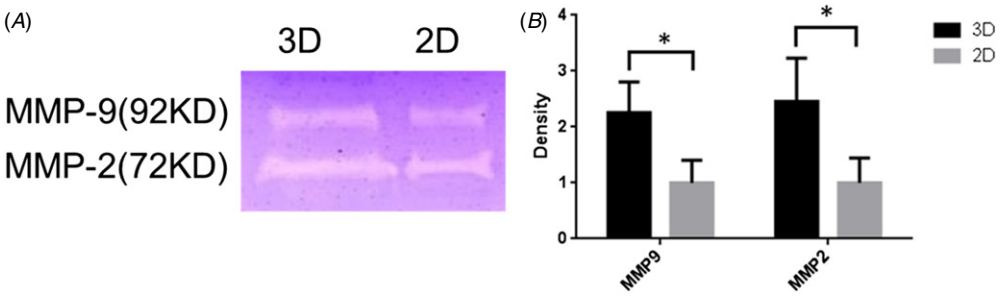

Figure 4. MMP secretion of Hela cells in 3D constructs and 2D planar culture. (*A*) MMP-9 and MMP-2 secretion of Hela cells in 3D constructs and 2D planar culture. (*B*) Semi-quantitative analysis of MMP-2 and MMP-9 secretion in 3D construct normalized to 2D samples. * means $p < 0.05$; *t*-test.

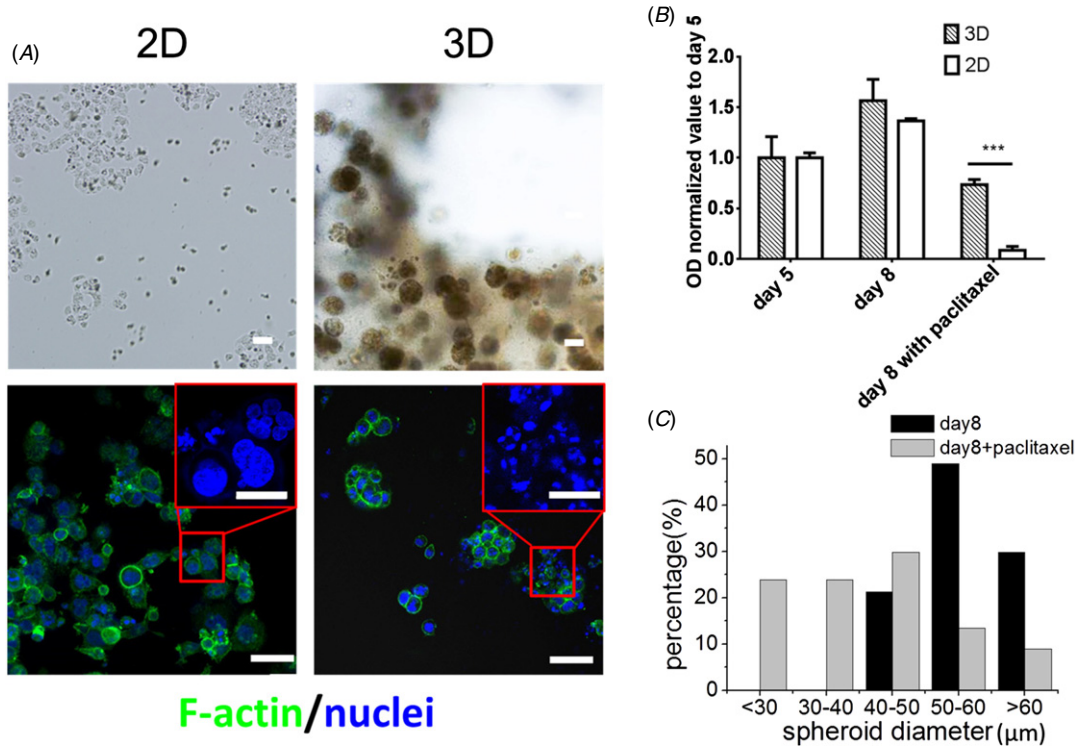

Figure 5. Chemoresistance of Hela cells in 3D Hela/hydrogel constructs and 2D planar culture. (*A*) Cell morphology after paclitaxel treatment in 2D planar culture and 3D Hela/hydrogel constructs. (*B*) Cellular metabolic activity after paclitaxel treatment in 2D planar culture and 3D Hela/hydrogel constructs. (*C*) Distribution of spheroid diameters in 3D Hela/hydrogel constructs on day 8 with and without the addition of paclitaxel. *** means $p < 0.001$; *t*-test. Scale bar, 50 μm (enlarged images, scale bar, 20 μm).

the viscosity of hydrogels results in an increase of shear forces on embedded cells, causing more cellular injury and death. As a thermosensitive hydrogel, the viscosity of gelatin increases with the decline of the hydrogel temperature. As a result, decreasing the chamber or nozzle temperature leads to increased hydrogel viscosity and decreased cellular viability after 3D printing. On the other hand, adequate hydrogel viscosity must be guaranteed to ensure the clear structure and stability of the 3D construct. In this study, we examined the process parameters to achieve both high cellular viability and a stable and clear structure (figures 1(*F*) and (*G*)).

Compared with the 2D planar culture, the additional dimensionality of 3D culture leads to differences in cell activities, including morphology, proliferation, and gene and protein expression [35]. We used an optical microscope and observed big differences in Hela morphology between 3D printed constructs and 2D planar culture. To further investigate this point, cytoskeletons and cell nuclei were treated by staining and observed under LSCM. Hela cells in 2D culture showed a monolayered morphology in the whole experimental period, whereas cellular spheroids were formed from single cells in 3D tumor-like constructs. Similar observations were reported for studies on epithelial cancer cells in 3D culture [36, 37]. Multicellular tumor spheroids with *in vivo* tumor characteristics of avascular tumor nodules are the classic 3D tumor models *in vitro* for anti-tumor drug testing [38–40]. However, the cellular spheroids established by traditional approaches (e.g. hanging drop [12]) were not embedded within matrix biomaterials. Cells were embedded within gels to promote cellular spheroid formation along with cell–matrix interactions [41]. Cellular spheroids in 3D gels usually have a polarized property that faces the gel at the basal compartment and encloses at the apical compartment [35], which is more similar to the manner *in vivo* compared with the 2D planar cell culture. The cell morphology in 3D printed Hela/hydrogel constructs was similar to that in 3D embedding gels [16]. We assumed that this was because we first mixed biomaterials (gelatin, alginate and fibrinogen) with single Hela cells and then printed the mixture layer by layer so that the cells were embedded within the hydrogels immediately after printing.

Enabling replicative immortality is one of the crucial cancer hallmarks [42], but tumor cell proliferation in 2D plates was inevitably inhibited by the area of the growth surface. Although Hela cells can form multi-layered growth without contact inhibition, the multi-layered cellular aggregates easily float away from the substrate and cause abundant cell loss. 10^6 mL^{-1} was the commonly used cell density in 3D construct fabrication [43]. When Hela cells were mixed with biomaterials at this density, 3D cellular spheroids were formed in 3D Hela/hydrogel constructs and continued growing until day 8 without obvious cell loss. However, when Hela cells were seeded at a similar density (10^6 cm^{-2}) as a 2D planar culture sample, they achieved 100% confluence in 2 to 3 days, and failed to complete the 8-day experiment. Finally, we chose to reduce the Hela cell seeding density to 5000 cells/cm^2 in 2D planar cell culture so as to prevent cell floating before 8 days of culture. This result also indicated an important advantage

of 3D cell/hydrogel constructs in supporting long-term cell proliferation and larger quantity number of cell delivery. We chose an 8 day experimental period in this study because cellular spheroids had occupied most of the area (79.5% ± 6.8%) in the matrix by day 8.

A CCK-8 kit was used for cell proliferation analysis. OD values of CCK-8 reagents were determined by the dehydrogenase activities in cells, which is in direct ratio to the number of living cells. In this study, we seeded the same number of cells in 2D plates (5000 cells/dish) and 3D constructs (5000 cells/construct) on day 0. However, OD values determined by a CCK-8 kit showed differences between 2D planar culture samples and 3D Hela/hydrogel constructs. We assumed that this was due to penetration differences of CCK-8 agents and/or cellular metabolic productions between elongated monolayered cells on 2D culture plates and spheroid single cells embedded in 3D hydrogels. The data of both the 3D and 2D samples were therefore normalized to day 0 to eliminate the influence of the proliferation kit. Hela cells in the 2D culture plates proliferated more slowly than in the printed 3D constructs with significant differences on day 5 and 8. We assumed that this was due to enhanced cell–cell interactions in the 3D cellular spheroids and cell–matrix interactions between the Hela and matrix biomaterials, although the detailed mechanisms need to be studied more closely. These results were also consistent with reported studies on 3D cervical tumor spheroid models [8].

The MMP protein family is considered fundamental in the degradation of the ECM [44, 45]. The MMP family, in particular MMP-2 and MMP-9, allow cancer cells to penetrate the ECM and are closely related to cancer metastasis [46]. The activity of MMPs tends to increase with the progression of cervical uterine neoplasms [47]. The study of cervical cancer *in vivo* confirmed a higher expression level of the MMP proteins in cervical tumor tissues compared with normal cervical tissues [48]. The activity of MMP proteins in cervical tumors was always studied *in vitro* based on Hela cells [49, 50]. We examined MMP-2 and MMP-9 expression levels as indicators of tumor metastasis characteristics of Hela cells in 2D and 3D conditions. Hela cells in 3D Hela/hydrogel constructs showed enhanced expression of MMP-2 as well as MMP-9 compared with 2D planar culture, indicating enhanced cellular metastasis in 3D printed constructs. This was similar to the studies on 3D glioma tumor models based on cell-seeding scaffolds [30].

Chemoresistance to anti-cancer drugs represents an important characteristic of enhanced tumor malignancy [51]. Monolayered cell culture with largely enhanced drug agent penetration always failed to mimic *in vivo* tumor characteristics. It has been demonstrated that cellular spheroids showed enhanced resistance to anti-tumor drugs compared with 2D planar cell culture [40, 52, 53]. Paclitaxel is a widely used anti-tumor drug which can mediate cell cycle arrest and cause apoptosis of tumor cells [16]. It is also known to induce a sustained mitotic block at the metaphase/anaphase boundary of Hela cells and inhibit Hela cell proliferation [3]. We observed massive cellular apoptosis both in 3D printed Hela/hydrogel constructs and 2D cell cultures after treatment with paclitaxel. After quantifying cellular metabolic activity with a CCK-8 kit, largely enhanced chemoresistance was observed in the 3D printed Hela/hydrogel constructs compared with the 2D planar cell culture, with significant differences. This result indicated the importance of dimensionality on the effectiveness of chemotherapy.

5. Conclusions

This paper reported a study of applying a 3D printing technique to construct *in vitro* cervical tumor models with Hela cells and gelatin/alginate/fibrinogen hydrogels. Cell proliferation, MMP protein expression and chemoresistance in the printed 3D cervical tumor models were measured and compared with the conventional 2D planar culture models. The study examined the effect of the printing parameters on cell viability, and a cell viability of over 90% was observed in the Hela cells under the printing process. Comparisons of 3D and 2D results reveal that the Hela cells showed a higher proliferation rate in the printed 3D environment and tended to form cellular spheroids, but formed monolayer cell sheets in the 2D culture. Hela cells in 3D printed models also showed higher MMP protein expression and higher chemoresistance than those in 2D culture. The results also reveal that the printed 3D models have more simulated tumor characteristics compared with the 2D planar cell culture models. Those 3D biological characteristics from the printed tumor models *in vitro* as well as the novel 3D cell printing technology may help the study of 3D tumor biology. In addition, the developed 3D printing process is capable of assembling cells with different phonotypes, thus allowing the construction of 3D *in vitro* models with heterogeneous cells to simulate the heterogeneous tumor microenvironment [54]. Therefore, the reported cell printing process may also have a broad application in the study of tumor heterogeneity [54].

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