

PAPER • OPEN ACCESS

Amplification and Analysis of Rbcl Gene (Ribulose-1,5-Bisphosphate Carboxylase) of Clove in Ternate Island

To cite this article: Nurhasanah *et al* 2019 *IOP Conf. Ser.: Earth Environ. Sci.* **276** 012061

View the [article online](#) for updates and enhancements.

You may also like

- [Green space in Ternate: tree species diversity and physical condition assessment](#)
R Ashari, L Irmayanti, Peniwidiyanti et al.
- [Population dynamics of eastern little tuna \(*Euthynnus affinis*\) in Ternate Waters](#)
U Tangke, R Laisouw, A Talib et al.
- [Sustainable practices: implementing bamboo cina conservation-based management in Ternate –North Maluku-Indonesia](#)
Mardiyani Sidayat



ECS
The
Electrochemical
Society
Advancing solid state &
electrochemical science & technology

DISCOVER
how sustainability
intersects with
electrochemistry & solid
state science research

Amplification and Analysis of RbcL Gene (Ribulose-1,5-Bisphosphate Carboxylase) of Clove in Ternate Island

Nurhasanah, Sundari*, Nurmaya Papuangan

Study Program of Biology Education, Faculty of Teacher Training and Education,
Khairun University Ternate, Indonesia

*Corresponding author: sundari@unkhair.ac.id

Abstract. DNA Barcoding is recommended as a tool for identifying and confirming species within the taxonomy framework. The rbcL gene is the barcode DNA for plant species. Ternate Island is one of clove plantation center in North Maluku. The diversity and productivity of cloves on the Ternate island known since in earlier times. This study has a purpose to amplify the rbcL gene in clove plants collected from clove plantations of communities in Ternate island. Isolation of total DNA carried out with using ZymoBiomic (Zymo Research DNA Extraction) Kit from leaves tissue, then DNA specimen is amplified based on rbcL gene with forwarding sequence rbcLaF 5'-ATG TCA CCA CAA ACA GAG ACT AAA GC-3' and reverse sequence is rbcLaR 5'-GTA AAA TCA AGT CCA CCR CG-3'. The results showed that the specimen was successfully amplified with an amplicon size of 600 bp. Furthermore, BLASTN analysis results note that the sequence has a similarity of 99% with *Syzygium cumini* vc. J.R. Abbott 23676 (FLAS) ribulose-1,5-bisphosphate carboxylase large subunit (rbcL) gene, but phylogenetically the precise position of sample could not found because the limitation of rbcL gene.

Keywords: Amplification, rbcL, clove, barcode DNA, Ternate

1. Introduction

North Maluku has the potential for spice plant diversity. Some of the world's spice plants are cloves and nutmeg. Clove commodities in North Maluku occupy the first rank so that the clove is an identity-flora. Ternate is one of the centers of clove plantations in North Maluku besides Tidore, Halmahera, Makian and Moti islands. Clove gardens range from small-scale communities to large-scale clove plantations spread across several of these islands. So far, there is no genetic information about the genetic diversity of clove varieties in Ternate island [1,2].

Genetic diversity in living things occurs through mutation and recombination mechanisms [3]. One approach used to assess genetic diversity is by DNA barcoding [4]. This method can be used to identify species [5] without having expert identification skills in the field [6]. DNA barcoding is usually used for pedigree reconstruction, forensics, and biodiversity surveys [7]. DNA barcoding is one of the methods in molecular level which has an approach to identify new species [8,9]. Last but not least, building a phylogenetic tree is not the main goal of DNA barcoding but to identify the unknown organism [10]. Chloroplast DNA (cpDNA) is a region highly conserved and often used in DNA barcodes for plants [11]. Chloroplast DNA (cpDNA) has a circular shape with a length between



85-2000 kilobase (kb); this region has a function to control the production of two types of RNA, i.e. tRNA and rRNA, also almost of proteins on chloroplasts organelles. Subunits complex form for photosynthetic protein contain with codes, and one of them is ribulose 1.5-biphosphate carboxylase oxygenase [12]. The cpDNA has several characters that are a stable structure, a small genome with a high conservative region and low substitution of nucleotide [13].

The *rbcL* gene is a part of DNA sequence located in cpDNA and has a chance to be used as a DNA barcode [14–17] because this coding region is given universality and ease in amplifying and analyzing [18]. This gene provides many characters to study phylogenetic because it has full length approximately 1400 bp [19]. This sequence has a low level of mutation compared with other barcodes in cpDNA, and because this sequence has a high level of similarity between species [20]. Low level of mutation is the superiority of the *rbcL* gene. Therefore, in-depth study of intraspecies genetic and phylogenetic variations can be done using this gene. This study aims to amplify the *rbcL* gene in clove plants collected from clove plantations in Ternate island community. Molecular phylogenetic data evidently can solve some taxonomic issues, where with the ways the data are hard to get through [21–26]. Based on the presented background, the research was conducted and the obtained results are expected to be used as a reference to develop cloves plant breeding programs on Ternate Island.

2. Methods

2.1. DNA isolation and amplification of *rbcL* gene

The leaves and stems of cloves plant were harvested from experimental plants to isolate the total DNA. The total DNA was isolated using DNA Presto TM Mini gDNA kit KIT (Geneid) kit. The PCR was proceeded by MyTag Red Mix (Bioline). Forward primer used in this research was *rbcLaF* (5'-ATG CCA CAA ACA GAG ACT AAA GC-3') and the reverse primer was *rbcLaR* (5'-GTA AAA TCA AGT CCA CCR CG-3') with total volume of PCR 30 mL. The PCR program was 95 °C for denaturation, 55 °C for annealing, and 72 °C for extension and 72 °C for final extension. Zimoclean TM gel DNA recovery KIT (Zimo research) used for purified PCR products. Sequencing was carried out in 1st Base Malaysia services.

2.2. Phylogenetic analysis

Some programs were used to analyze the obtained data. MEGA5 was employed for DNA alignment and also construction of the Phylogenetic tree with Neighbor Joint (NJ) method; BLASTn was utilize to compare the obtained sequence with DNA sequences from GenBank.

3. Results and Discussion

DNA barcoding is one of the ways to fulfill The Barcode of Life Database which aims to collect the reference sequences. This effort used variations of shorts standardized gene regions to identify new species [8]. The first step in DNA barcoding was the isolation of total DNA from the sample. The total DNA of cloves plant was successfully obtained. The next step was measuring the quality and quantity of DNA using DNA spectrophotometer and agarose electrophoresis. The purity and concentration of the obtained DNA are presented in Table 1.

Table 1. The results of DNA isolation

No	Sample name	Conc (ng/mL)	A260/280	A260/230	Volume (µL)
1	MB1	37	1.83	0.17	30
2	SD1	21.7	1.92	0.32	30

The table shows the two samples of DNA isolation of clove plants had 37 ng/µL and 21.7 ng/µL with a purity between 1.8-2.0. The next step was the amplification of DNA using the *rbcL* gene. Genome DNA containing the *rbcL* gene was amplified by polymerase chain reaction (PCR). The

amplification of the *rbcL* gene that had been successfully carried out was tested by electrophoresis (Figure 1).

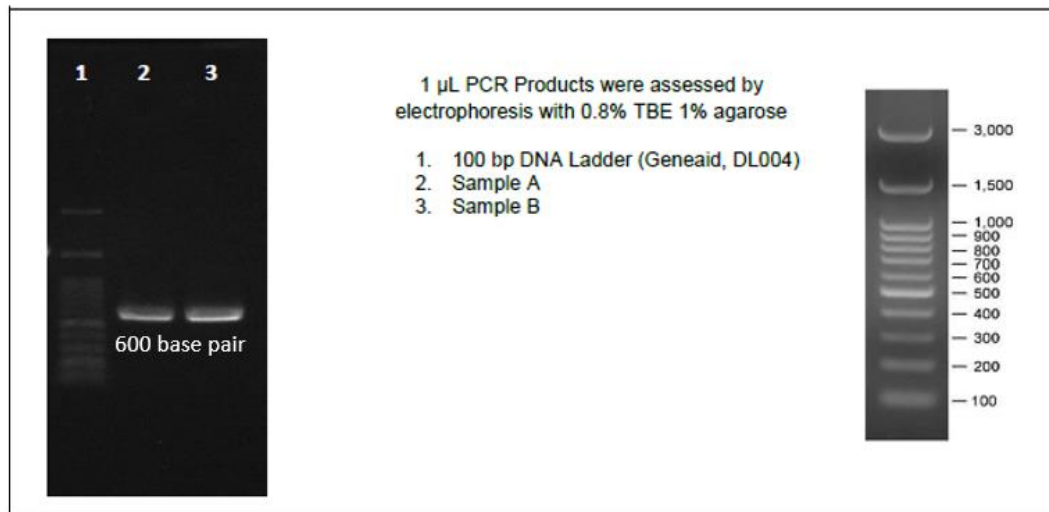


Figure 1. Electropherogram results of the *rbcL* gene amplification in clove plants from Ternate

DNA band obtained from the amplification of the *rbcL* gene was ± 600 bp. The sample of SD1 primer *rbcL* does not proceed to sequence because it cannot be amplified even though PCR repeats have been performed. Furthermore, the BLASTN analysis showed that MB 1 isolates were identical to *Syzygium cumini* J.R. vc. Abbott 23676 (FLAS) ribulose-1,5-bisphosphate carboxylase/oxygenase large subunit (*rbcL*) gene, with an identical value of 99%. The data from the similarity analysis (Blastn) from the MB1 sample are shown in Figure 2 below:

	Description	Max score	Total score	Query cover	E value	Ident	Accession
✓	Luma apiculata voucher OMH-A58 ribulose-1,5-bisphosphate carboxylase/oxygenase large subunit (rbcL) gene, partial cds: chloroplast	908	908	100%	0.0	99%	KX162972.1
✓	Backhousia citriodora ribulose-1,5-bisphosphate carboxylase/oxygenase large subunit gene, partial cds: chloroplast	904	904	100%	0.0	99%	JN676920.1
✓	Acca sellowiana chloroplast, complete genome	902	902	100%	0.0	99%	KX289887.1
✓	Uromyrtus australis ribulose-1,5-bisphosphate carboxylase/oxygenase, large subunit (rbcL) gene, partial cds: chloroplast	902	902	100%	0.0	99%	KU761898.1
✓	Campomanesia xanthocarpa voucher UPCB:UFPR Blum10-109 ribulose-1,5-bisphosphate carboxylase/oxygenase large subunit (rbcL) gene, partial cds: chloroplast	902	902	100%	0.0	99%	KF561906.1
✓	Campomanesia xanthocarpa voucher UPCB:UFPR Blum10-101 ribulose-1,5-bisphosphate carboxylase/oxygenase large subunit (rbcL) gene, partial cds: chloroplast	902	902	100%	0.0	99%	KF561905.1
✓	Campomanesia xanthocarpa voucher UPCB:UFPR Blum10-100 ribulose-1,5-bisphosphate carboxylase/oxygenase large subunit (rbcL) gene, partial cds: chloroplast	902	902	100%	0.0	99%	KF561904.1
✓	Syzygium cumini chloroplast, complete genome	902	902	100%	0.0	99%	GQ870669.3
✓	Melaleuca quinquenervia voucher J.R. Abbott 23686 (FLAS) ribulose-1,5-bisphosphate carboxylase/oxygenase large subunit (rbcL) gene, partial cds: chloroplast	902	902	100%	0.0	99%	GU135164.1
✓	Syzygium cumini voucher J.R. Abbott 23676 (FLAS) ribulose-1,5-bisphosphate carboxylase/oxygenase large subunit (rbcL) gene, partial cds: chloroplast	902	902	100%	0.0	99%	GU135161.1

Figure 2. The analysis of BLAST *rbcL* sequence of clove plants

Phylogenetic analysis was carried out to find the species name of the sample. The first step was comparing the sequence of the sample with the data from GenBank using BLASTn method in NCBI. The position in taxon from clove samples could be known from this analysis. The result of the relationship analysis (phylogenetic) of clove samples is presented in Figure 3 below:

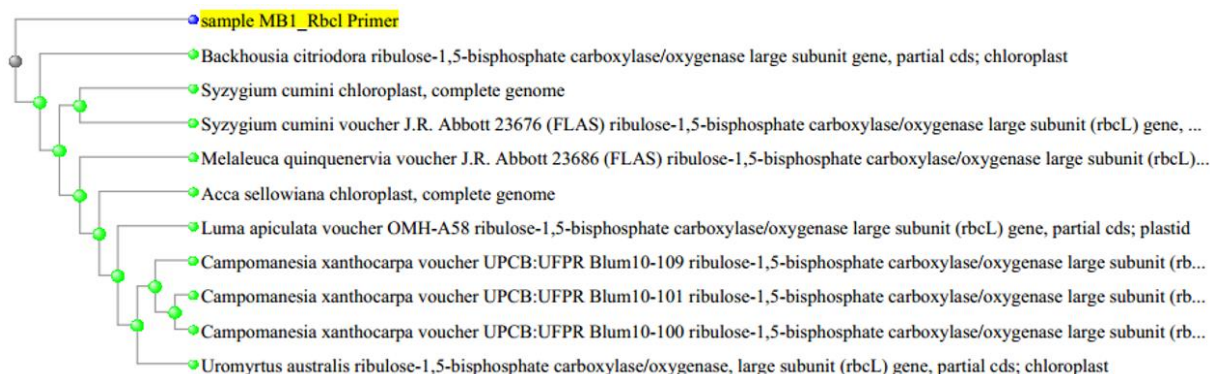


Figure 3: Phylogenetic analysis from a sample of MB 1 Ternate clove plants

Neighbor-Joining (NJ) method in the analysis of phylogenetic can describe the clarity of species identification; the difference is limited by cluster and node. The Sample can be in the same cluster even though they are from different areas [7]. Species relationship based on genetic similarities is shown in the phylogenetic tree. The samples of MB1 clove plants were located in different clusters with *Synzigium cumini* (Figure 3). The results of BLASTn analysis showed the *rbcL* gene clove sample with MB 1 code which had a 99% similarity with *Syzygium cumini* J.R vc. Abbott 23676 showed a distant taxon position. The databases on the *rbcL* gene have been owned by many species, making it easier to compare data analysis [28].

Only MB 1 sample of clove in Ternate that was successful in amplifying using *rbcL* gene. Meanwhile, the DNA band of the *rbcL* gene did not appear on SD 1 sample; this condition might occur because the sample was degraded. The amplification of the *rbcL* gene using one or two universal primer types had a high success rate [19]. Moreover, using the *rbcL* gene rate of success bidirectional (two-way sequencing with forward and reverse primers) sequencing increase higher compared with other barcode gene candidates. The success rate of the *rbcL* gene could reach 100% in 251 plant species with only two primary types [29]. Another researcher recommended using four primers for *rbcL* has a significantly greater length (approximately 1.428 bp) [30]. However, the *rbcL* gene is essential to identify phylogenetic relationship at interfamilial level [31].

cpDNA coded many proteins, and one of them is Ribulose-1,5-biphosphate carboxylase-oxygenase (RuBisCO) which has participated in carbon fixation in photosynthesis [32]. This enzyme contains two kinds of protein subunit called the large chain and the small chain. The large chain gene (*rbcL*) is encoded by cpDNA and has been widely used for analysis of phylogenetic in plant taxonomy [16]. The utilization of the *rbcL* gene to identify in species level was rejected [31,33,34]. The phylogenetic based on the *rbcL* gene is usually used at the generic or higher level or [30,31]. This case was proven in the previous study that used the *rbcL* gene to reveal monophyly in the order of Myrtales [35,36]. It is suggested to use non-coding regions of cpDNA like *tnhpsb5* and *ndITs* to analyze the phylogenetic flowering plants at lower taxonomic levels, not only because this region tends to evolve rapidly rather than coding sequence gene but also this region has a smaller length and thus phylogenetic could be easier to analyze [10,30,31]. Non-coding DNA has a higher number of variable sites when compared with coding DNA, because Non-coding DNA is better to be used in phylogenetic [37] and the single region in the plastid genome do not have variable sequence enough to be used as a barcode gene [38]. Furthermore, the study compared between *ndhF*, *matK*, and *rbcL* gene showing both *ndhF* and *matK* faster-evolving rather than *rbcL* [39]. Another option besides using non-coding regions in phylogenetic is using combination between *rbcL* + *matK*, because *rbcL* gene is easy in amplifying and analyzing [40] while *matK* is coding sequence located in plastid genome and has a high rate to evolving [41]. This combination performs slightly higher than *rbcL* alone [6].

Comparing the divergence of plastid genomes in *Atropa* and *Nicotina*, resulted in the lowest divergence in plastid genome of *rbcL* gene (0.83%) [30]. Meanwhile, another study compared the species discrimination between 7 leading candidate plastid DNA regions; the result was *rbcL* gene had

58% - 66% of single-locus barcodes range [19]. In addition, another study also compared the use of *rbcL* gene to identify genus and species level, the results showed the percentage of correct identify in genus level was 67.71% and in species level is 16.95% [18]. Therefore, the *rbcL* gene had been knocked off for discrimination in the level of species [30].

4. Conclusion

The *rbcL* gene that had been successfully amplified from one sample of clove plant in this study, had a length of 600 bp. The sequence had a similarity of 99% with JR's voucher *Syzygium cumini*. The exact species could not be found because the *rbcL* gene could be identified at the generic level or higher.

References

- [1] Arifin H S, Sardjono M A, Sundawati L and Djogo T 2003 Agroforestri di Indonesia 90
- [2] CGIAR T A M 1988 Sustainable Agricultural Production: Implications for International Agricultural Research
- [3] Indrawan M, Primack R B and Supriatna 2012 *Biologi Konservasi (Edisi Revisi)* (Jakarta: Yayasan Pustaka Obor Indonesia)
- [4] Blaxter M L 2004 The promise of a DNA taxonomy ed H C J Godfray and S Knapp *Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences* **359** 669–79
- [5] Shen Y-Y, Chen X and Murphy R W 2013 Assessing DNA Barcoding as a Tool for Species Identification and Data Quality Control ed D Steinke *PLoS ONE* **8** e57125
- [6] Costion C, Ford A, Cross H, Crayn D, Harrington M and Lowe A 2011 Plant DNA Barcodes Can Accurately Estimate Species Richness in Poorly Known Floras ed J A Gilbert *PLoS ONE* **6** e26841
- [7] Che J, Chen H-M, Yang J-X, Jin J-Q, Jiang K, Yuan Z-Y, Murphy R W and Zhang Y-P 2012 Universal COI primers for DNA barcoding amphibians: UNIVERSAL COI PRIMERS FOR DNA BARCODING AMPHIBIANS *Molecular Ecology Resources* **12** 247–58
- [8] Gutteridge A and Burns M 2013 The Application of DNA Molecular Approaches for the Identification of Herbal Medicinal Products *Journal of the Association of Public Analysts* 14
- [9] Park D-S, Suh S-J, Oh H-W and Hebert P D 2010 RReeseacrcohvareticrley of the mitochondrial COI barcode region in diverse Hexapoda through tRNA-based primers 7
- [10] Kress W J and Erickson D L 2007 A Two-Locus Global DNA Barcode for Land Plants: The Coding *rbcL* Gene Complements the Non-Coding *trnH-psbA* Spacer Region ed S-H Shiu *PLoS ONE* **2** e508
- [11] Asif H, Khan A, Iqbal A, Khan I A, Heinze B and Azim M K 2013 The chloroplast genome sequence of *Syzygium cumini* (L.) and its relationship with other angiosperms *Tree Genetics & Genomes* **9** 867–77
- [12] Tamarin R H 2001 *Principles of Genetics* (Massachusetts: The McGraw-Hill Companies)
- [13] Provan J, Powell W and Hollingsworth P M 2001 Chloroplast microsatellites: new tools for studies in plant ecology and evolution *Trends in Ecology & Evolution* **16** 142–7
- [14] Les D H, Garvin D K and Wimpee C F 1991 Molecular evolutionary history of ancient aquatic angiosperms. *Proceedings of the National Academy of Sciences* **88** 10119–23
- [15] Duvall M R, Clegg M T, Chase M W, Clark W D, Kress W J, Hills H G, Eguiarte L E, Smith J F, Gaut B S, Zimmer E A and Learn G H 1993 Phylogenetic Hypotheses for the Monocotyledons Constructed from *rbcL* Sequence Data *Annals of the Missouri Botanical Garden* **80** 607
- [16] Chase M W, Soltis D E, Olmstead R G, Morgan D, Les D H, Mishler B D, Duvall M R, Price R A, Hills H G, Qiu Y-L, Kron K A, Rettig J H, Conti E, Palmer J D, Manhart J R, Sytsma K J, Michaels H J, Kress W J, Karol K G, Clark W D, Hedren M, Gaut B S, Jansen R K, Kim K-J, Wimpee C F, Smith J F, Furnier G R, Strauss S H, Xiang Q-Y, Plunkett G M, Soltis P S, Swensen S M, Williams S E, Gadek P A, Quinn C J, Eguiarte L E, Golenberg E, Learn G H,

- Graham S W, Barrett S C H, Dayanandan S and Albert V A 1993 Phylogenetics of Seed Plants: An Analysis of Nucleotide Sequences from the Plastid Gene *rbcL* *Annals of the Missouri Botanical Garden* **80** 528
- [17] Hasebe M, Omori T, Nakazawa M, Sano T, Kato M and Iwatsuki K 1994 *rbcL* gene sequences provide evidence for the evolutionary lineages of leptosporangiate ferns. *Proceedings of the National Academy of Sciences* **91** 5730–4
- [18] Newmaster S G, Fazekas A J and Ragupathy S 2006 DNA barcoding in land plants: evaluation of *rbcL* in a multigene tiered approach *Canadian Journal of Botany* **84** 335–41
- [19] CBOL Plant Working Group, Hollingsworth P M, Forrest L L, Spouge J L, Hajibabaei M, Ratnasingham S, van der Bank M, Chase M W, Cowan R S, Erickson D L, Fazekas A J, Graham S W, James K E, Kim K-J, Kress W J, Schneider H, van AlphenStahl J, Barrett S C H, van den Berg C, Bogarin D, Burgess K S, Cameron K M, Carine M, Chacon J, Clark A, Clarkson J J, Conrad F, Devey D S, Ford C S, Hedderson T A J, Hollingsworth M L, Husband B C, Kelly L J, Kesanakurti P R, Kim J S, Kim Y-D, Lahaye R, Lee H-L, Long D G, Madrinan S, Maurin O, Meusnier I, Newmaster S G, Park C-W, Percy D M, Petersen G, Richardson J E, Salazar G A, Savolainen V, Seberg O, Wilkinson M J, Yi D-K and Little D P 2009 A DNA barcode for land plants *Proceedings of the National Academy of Sciences* **106** 12794–7
- [20] Kellogg E A and Juliano N D 1997 The structure and function of RuBisCO and their Implications for Systematic Studies *American Journal of Botany* **84** 413–28
- [21] O'Brien M M, Quinn C J and Wilson P G 2000 Molecular systematics of the Leptospermum suballiance (Myrtaceae) *Australian Journal of Botany* **48** 621
- [22] Brown G K, Udovicic F and Ladiges P Y 2001 Molecular phylogeny and biogeography of Melaleuca, Callistemon and related genera (Myrtaceae) *Australian Systematic Botany* **14** 565
- [23] Wilson P G, O'Brien M M, Gadek P A and Quinn C J 2001 Myrtaceae revisited: a reassessment of infrafamilial groups *American Journal of Botany* **88** 2013–25
- [24] Wright S D, Yong C G, Wichman S R, Dawson J W and Gardner R C 2003 Stepping stones to Hawaii: a trans-equatorial dispersal pathway for Metrosideros (Myrtaceae) inferred from nrDNA (ITS+ETS): Stepping stones to Hawaii *Journal of Biogeography* **28** 769–74
- [25] Harrington M G and Gadek P A 2004 Molecular systematics of the Acmena alliance (Myrtaceae): phylogenetic analyses and evolutionary implications with reference to Australian taxa *Australian Systematic Botany* **17** 63
- [26] Biffin E, Craven L A, Crisp M D and Gadek P A 2006 Molecular systematics of *Syzygium* and allied genera (Myrtaceae): evidence from the chloroplast genome *TAXON* **55** 79–94
- [27] Tamura K, Peterson D, Peterson N, Stecher G, Nei M and Kumar S 2011 MEGA5: Molecular Evolutionary Genetics Analysis Using Maximum Likelihood, Evolutionary Distance, and Maximum Parsimony Methods *Molecular Biology and Evolution* **28** 2731–9
- [28] Cummings M P, Nugent J M, Olmstead R G and Palmer J D 1994 Phylogenetic analysis reveals five independent transfers of the chloroplast gene *rbcL* to the mitochondrial genome in angiosperms **8**
- [29] Fazekas A J, Burgess K S, Kesanakurti P R, Graham S W, Newmaster S G, Husband B C, Percy D M, Hajibabaei M and Barrett S C H 2008 Multiple Multilocus DNA Barcodes from the Plastid Genome Discriminate Plant Species Equally Well ed R DeSalle *PLoS ONE* **3** e2802
- [30] Kress W J, Wurdack K J, Zimmer E A, Weigt L A and Janzen D H 2005 Use of DNA barcodes to identify flowering plants *Proceedings of the National Academy of Sciences* **102** 8369–74
- [31] Gielly L and Taberlet P 1994 The use of chloroplast DNA to resolve plant phylogenies: noncoding versus *rbcL* sequences. *Mol Biol Evol* **11** 769–77
- [32] Pierce B A 2002 *Genetics: A Conceptual Approach* (New York: W. H. Freeman and Company)
- [33] Renner S S 1999 Circumscription and phylogeny of the Laurales: evidence from molecular and morphological data *American Journal of Botany* **86** 1301–15

- [34] Salazar G A, Chase M W, Soto Arenas M A and Ingrouille M 2003 Phylogenetics of Cranichideae with emphasis on Spiranthinae (Orchidaceae, Orchidoideae): evidence from plastid and nuclear DNA sequences *American Journal of Botany* **90** 777–95
- [35] Conti E, Litt A and Sytsma K J 1996 Circumscription of Myrtales and their Relationships to Other Rosids: Evidence from rbcL Sequence Data *American Journal of Botany* **83** 221
- [36] Conti E, Litt A, Wilson P G, Graham S A, Briggs B G, Johnson L A S and Sytsma K J 1997 Interfamilial Relationships in Myrtales: Molecular Phylogeny and Patterns of Morphological Evolution *Systematic Botany* **22** 629
- [37] Borsch T and Quandt D 2009 Mutational dynamics and phylogenetic utility of noncoding chloroplast DNA *Plant Systematics and Evolution* **282** 169–99
- [38] Chase M W, Salamin N, Wilkinson M, Dunwell J M, Kesanakurthi R P, Haidar N and Savolainen V 2005 Land plants and DNA barcodes: short-term and long-term goals *Philosophical Transactions of the Royal Society B: Biological Sciences* **360** 1889–95
- [39] Sytsma K J, Litt A, Zjhra M L, Chris Pires J, Nepokroeff M, Conti E, Walker J and Wilson P G 2004 Clades, Clocks, and Continents: Historical and Biogeographical Analysis of Myrtaceae, Vochysiaceae, and Relatives in the Southern Hemisphere *International Journal of Plant Sciences* **165** S85–105
- [40] Hollingsworth P M, Graham S W and Little D P 2011 Choosing and Using a Plant DNA Barcode ed D Steinke *PLoS ONE* **6** e19254
- [41] Hilu K W and Liang gping 1997 The matK gene: sequence variation and application in plant systematics *American Journal of Botany* **84** 830–9