PAPER • OPEN ACCESS

Detection of antibiotic-resistant *Salmonella* sp. in the seafood products of Surabaya local market

To cite this article: H Pramono et al 2019 IOP Conf. Ser.: Earth Environ. Sci. 236 012115

View the article online for updates and enhancements.

You may also like

- Occurrences of Salmonella spp. and Escherichia coli in chicken meat, intestinal contents and rinse water at slaughtering place from traditional market in Surabaya, Indonesia

R Yulistiani, D Praseptiangga, Supyani et al.

- <u>Thermal Stability of Phage Peptide Probes</u> <u>Vs. Aptamer for Salmonella Detection on</u> <u>Magnetoelastic Biosensors Platform</u> I-Hsuan Chen, Shin Horikawa, Songtao Du et al.
- <u>Real-Time Bacterial Detection in Liquid By</u> <u>Using Magnetoelastic Biosensors and a</u> <u>Surface-Scanning Coil Detector</u> Songtao Du, Yuzhe Liu, Shin Horikawa et al.





DISCOVER how sustainability intersects with electrochemistry & solid state science research



This content was downloaded from IP address 18.218.234.83 on 01/05/2024 at 21:04

Detection of antibiotic-resistant Salmonella sp. in the seafood

products of Surabaya local market

H Pramono^{1*}, A Kurniawan², N Andika², T F Putra², M A R Hazwin², S Utari², A P Kurniawan², E D Masithah¹ and A M Sahidu¹

¹ Department of Marine, Faculty of Fisheries and Marine, UniversitasAirlangga, Campus C UNAIR, Jl Mulyorejo, Surabaya, Indonesia 60115

² Aquaculture Study Program, Faculty of Fisheries and Marine, UniversitasAirlangga Campus C UNAIR, Jl Mulyorejo, Surabaya, Indonesia 60115

*Corresponding author: heru.pramono@fpk.unair.ac.id

Abstract. The consumption of seafood and seafood products contaminated with *Salmonella* sp. can lead to illness, thus this has become a worldwide public health concern. The aim of this research was to detect Salmonella sp. in various seafood and seafood products from Surabaya traditional market. The study was conducted through a sample collection, followed by the preenrichment, enrichment, isolation, and the identification of Salmonella. A total of twenty-nine samples (fish, crustacean, bivalve, and seafood products) from five local markets in Surabaya were collected and analyzed. Twenty-seven (93.10%) samples were positive for Salmonella sp. All of the isolates were tested for an antibiotic-resistant profile by employing disc diffusion targeting seven antibiotics. A high prevalence of Salmonella sp. in the seafood in the traditional markets was apparent in this study. Among all of the isolates, there were eight strains resistant to at least one antibiotic that was tested. This study can be a basis for further research into managing antibiotic-resistant Salmonella sp. infection as an important food-borne illness of developing countries.

1. Introduction

Salmonella sp. is normally found in humans and animal digestive tracts. However, it is also widespread in the environment, can be consumed by humans and can cause illness [1]. The bacterium can reach the human gastrointestinal tract through the consumption of contaminated foods such as eggs, milk, poultry, meat, aquaculture products and seafood [2]. Salmonella naturally cannot be found in seafood. However, the contamination of it during handling and processing can be a serious problem for public health [3].

In recent years, many articles have reported the occurrence of Salmonella in fish products. Raufu et al. [2] said that 23 (11.5%) out of 200 fish samples were Salmonella positive in sub-Saharan Africa. In another study, Zhang et al. [3] reported that 217 (29.7%) out of 730 fishery products were Salmonella positive. On the other hand, many reports were focused on the occurrence of Salmonella in terrestrial animal products, such as raw eggs, raw meat, mayonnaise, turkey meat, broiler chicken and sausage [1]. Therefore, the occurrence of *Salmonella* in seafood and seafood products is a recent advance in detection as part of an overall mitigation strategy.

Human health surveillance programs are lacking in coastal communities due to their limited access to transport-wise and poor personal hygiene. On the other hand, many coastal areas of Indonesia are

Content from this work may be used under the terms of the Creative Commons Attribution 3.0 licence. Any further distribution of this work must maintain attribution to the author(s) and the title of the work, journal citation and DOI. Published under licence by IOP Publishing Ltd 1

becoming centers of human activity, such as Jakarta and Surabaya. Surabaya, the second biggest city in Indonesia, is located in the East Java Province. A common part of the daily consumption of food by Surabaya's people is the seafood that can be obtained from fishermen, traditional markets or supermarkets. Thus, the data on food-borne pathogens related to seafood and seafood products have become important as part of a wider mitigation strategy. However, up until now, the data on the occurrence of *Salmonella* in seafood and seafood products is still limited.

Focusing on food-borne pathogens, particularly antibiotic-resistant bacteria, it is a major worldwide concern. Infected patients will need special treatment to be cured. Food-borne illnesses consist of *Salmonella, Campylobacter, Escherichia coli*, and *Listeria monocytogenes* [4]. Therefore, this study focused on the antibiotic-resistant profile of *Salmonella* is important.

The occurrence of *Salmonella* in fresh shrimp in five traditional markets in Surabaya reported a rate of occurrence of 36%. On the other hand, the data for *Salmonella* in other seafood (fish, cephalopods, shellfish, and seafood products) has not yet been reported. Therefore, the objective of this study was to determine the occurrence of antibiotic-resistant *Salmonella* in seafood and seafood products in Surabaya, East Java, Indonesia.

2. Materials and methods

2.1. The study area

The study area, Surabaya, located in East Java, is the second biggest city in Indonesia and is populated by 2,000,000 people. The sampling locations chosen consisted of traditional west, north, east, south and central Surabaya local markets selling seafood and seafood products.

2.2. Sample collection

A total of 29 samples (17 fish, 4 crustacea, 5 mollusks, and 3 fermented fish products) were collected from five traditional fish markets in Surabaya and analyzed for the presence of *Salmonella*. All of the samples were collected using a standard aseptic protocol and transported to the Education Laboratory of Faculty of Fisheries and Marine, Universitas Airlangga. The samples were analyzed directly upon arrival at the laboratory.

2.3. Salmonella isolation

For the isolation, 25 g of seafood sample was homogenized with 225 ml of peptone water and then pre-enriched at 37°C for 18 h. The culture was taken from the pre-enrichment broth (1 ml) and then sub-cultivated in two selective enrichment broths; a Selenite Cystine Broth (Merck) and a Rappaport-Vassiliadis Broth (Oxoid). The broths were incubated for 24 h at 41°C. A loopful sample from the enrichment broths was streaked on the selective plating media *Salmonella* Shigella Agar (Merck) and Xylose Lysine Deoxycholate (Merck) and incubated at 37 C for 24 h. The *Salmonella* presumptive colonies that showed a clear black-centered colony were taken for further purification steps. The next purification step was done by streaking the *Salmonella* presumptive colonies onto tryptic soy agar (Merck) incubated at 37°C for 24 h. A single colony with demonstrated growth was then taken for Gram staining and biochemical testing for confirmation.

2.4. Profiling of the Antibiotic-Resistant Salmonella

An antibiotic-resistant test was conducted by Kirby-Bauer disc diffusion assay against seven antibiotics: chloramphenicol (30 μ g), lincomycin (10 μ g), azithromycin (15 μ g), cefixime (5 μ g), amoxicillin, trimethoprim/sulfamethoxazole (1.25/23.75 μ g), erythromycin (15 μ g), ciprofloxacin (5 μ g), tetracycline (30 μ g) and ampicillin (10 μ g). Briefly, one loopful of purified Salmonella was cultured in tryptic soy broth (TSB) overnight, and the cell was separated from the broth through a centrifuge (Hettich Centrifuge, Germany) at 5,000 rpm for 5 minutes and then washed with normal saline twice. The cell was adjusted to McFarland standard number 1 and then streaked on Mueller Hinton Agar (Merck) with a sterile cotton butt. Antibiotic discs were placed on the lawn and incubated

IOP Conf. Series: Earth and Environmental Science 236 (2019) 012115 doi:10.1088/1755-1315/236/1/012115

at 4°C for 1 h prior to incubation at 37°C for 24 h. The clear zone was measured and the resistance strain was decided by CLSI [5].

3. Results and discussion

3.1. Samples profile

A total of 29 samples were obtained from 5 traditional markets in Surabaya, consisting of fish, crab, shrimp, shrimp products and cuttlefish. Details of the traditional markets as a source of the samples have been shown in Table 1.

Table 1. Details of the seafood and seafood product samples.								
	Fish a	ind	Shrimp an	Shrimp and				
Traditional Market	fish	Crab	shrimp	Cuttlefish	Shellfish			
	products		products	products				
Babatan/Wiyung	10	-	2	1	1			
Sutorejo	5	-	2	-	1			
Kenjeran	1	-	-	-	1			
Mulyorejo	-	-	1	-	-			
Waru	1	1	1	-	1			

3.2. Occurrence of Salmonella

Traditional markets or retail markets have become one site of *Salmonella* contamination that can affect public health concerns [6]. A total of 27 isolates out of 29 seafood and seafood product samples were found to be *Salmonella* positive based on a culture analysis. This represents a prevalence of 93%. The details of the data have been shown in Table 2.

Products	Number of positive samples/total number of samples	Percentage (%)
Fish	16/17	94,11
Crab	1/1	100
Shrimp and shrimp product	6/6	100
Cuttlefish	1/1	100
Shellfish	3⁄4	75

Table 2. Salmonella prevalence in various seafood and seafood products in Surabaya.

In our study, the high prevalence of *Salmonella* in the fish samples (94%) was in agreement with [7], which reported that the prevalence of *Salmonella* in the fish samples was 90%. However, relatively lower results were reported by [3], [8] and [6] at rates of 29.7%, 43.8%, and 5% respectively. The high prevalence of shrimp in this study (100%) and shrimp products (100%) was also with an agreement with [9], at a rate of 58%. However, Yang et al. [10] reported a relatively low rate of *Salmonella* in the shrimp products (13.0%). The high prevalence of *Salmonella* in the shrimp products may occur due to *Salmonella* being a part of the cultural environment and natural flora [11]. On the other hand, the high rate of *Salmonella* in cephalopod products from India as 3.2%. The difference in the results may be caused by several factors, including sanitation, the isolation procedure and the sampling season [13].

Our finding in this study was that there is a high prevalence of Salmonella in the seafood and seafood products in Surabaya's traditional markets. Gastroenteritis caused by Salmonella sp. is considered to be a worldwide public health problem and the first step to overcoming the problem is understanding the Salmonella prevalence, cross-contamination process and personal hygiene. Moreover, antibiotic resistance to Salmonella has also become a worldwide concern [2,3,6].

Therefore, further research related to antibiotic resistance profiling on Salmonella isolated from Surabaya's traditional markets is needed.

3.3. Antibiotic-resistant profile

Among the 33 isolates tested, there were eight isolates resistant to antibiotics (Table 3).

No Code	Products	Antibiotics											
		1	2	3	4	5	6	7	8	9	10		
1	SBI04	Pindang	S	R	S	S	S	S	R	S	R	S	
2	SBI07	IkanCukul	S	R	R	S	S	S	R	S	S	S	
3	SBK01	Anadara	R	R	S	S	S	S	R	S	S	S	
4	SBI01	Belanak	S	R	S	S	S	S	R	S	R	S	
5	SSI01	Banyar	S	R	S	S	S	S	R	S	R	S	
6	SSP01	Kepiting	S	R	S	S	S	S	R	S	S	S	
7	STI02	Jambal	S	R	R	S	S	S	R	S	R	S	
8	STI04	Ikanlaut	R	R	R	S	S	S	R	S	S	S	

 Table 3. Resistance profile of Salmonella strains isolated from the seafood of Surabaya Local Market.

Note: R (Resistant), S (Susceptible), 1. Chloramphenicol (30 μg); 2. lincomycin (10 μg); 3. azithromycin (15 μg); 4. Cefixime (5 μg); 5. Amoxicillin; 6. Trimethoprim/sulfamethoxazole (1.25/23.75 μg); 7. Erythromycin (15 μg); 8. Ciproflaxin (5 μg); 9. Tetracyclin (30 μg) and 10. Ampicillin (10 μg)

The resistance test results showed that 12.12% of the isolates were resistant to tetracycline and that 100% were resistant to erythromycin. These results are in contrast to the studies conducted by Nguyen et al. [14] that obtained the result that 53.3% were resistant to tetracycline, 43.8% resistant to ampicillin, chloramphenicol 37.5%, and 31.3% for trimethoprim/sulfamethoxazole. The testing of the antibiotic resistance of the Salmonella isolates also showed that there were traditional markets with samples that were resistant to antibiotics. Resistance to antibiotics is one of the world's focuses on attention because if the antibiotic-resistance test results showed that the majority of the isolates were sensitive to the antibiotics used; however, there were 8 isolates that were resistant to at least one antibiotic, and even resistant to more than two antibiotics (multidrug resistant).

4. References

- [1] Kramarenko T, Nurmoja I, Kärssin A, Meremäe K, Hörman A and Roasto M 2014 Food Control **1** 43-47
- [2] Raufu I A, Lawan F A, Bello H S, Musa A S, Ameh J A and Ambali A G 2014 *The Egyp. J. Aquat. Res* **40** 59-63
- [3] Zhang J, Yang X, Kuang D, Shi X, Xiao W, Zhang J, Gu Z, Xu X and Meng J 2015 Int. J. Food Microbiol. 210 47-52
- [4] Newell D G, Koopmans M, Verhoef L, Duizer E, Aidara-Kane A, Sprong H, Opsteegh M, Langelaar M, Threfall J, and Scheutz F 2010 *Inter. J. Food Microbiol.* **30**, 3-15
- [5] Cockerill F R, Wikler M A, Bush K, Dudley M N, Eliopoulus G M, Hardy D J, Hecht D W, Hindler J A, Patel J B, Powell M et al. 2010 *Performance Standards for Antimicrobial Susceptibility Testing* (USA: Clinical and Laboratory Standards Institute) p 172
- [6] Onmaz N E, Abay S, Karadal F, Hizlisoy H, Telli N and Al S 2015 *Mar. Pollut. Bull.* **15** 242-246
- [7] Jegadeeshkumar D, Saritha V, Moorthy K and Suresh K B T 2010 Int. J. Biol. Tech. 1 50-5
- [8] Budiati T, Rusul G, Wan-Abdullah W N, Arip Y M, Ahmad R and Thong K L 2013 Aquaculture 24 127-132
- [9] Allshouse J E, Buzby J, Harvey D and Zorn D 2004 *Seafood Safety and Trade* (USA: Department of Agriculture, Economic Research Service)

IOP Conf. Series: Earth and Environmental Science 236 (2019) 012115 doi:10.1088/1755-1315/236/1/012115

- [10] Yang X, Wu Q, Zhang J, Huang J, Chen L, Liu S, Yu S and Cai S 2015 Food Control 57 308-413
- [11] Bhaskar N, Setty T R, Reddy G V, Manoj Y B, Anantha C S, Raghunath B S and Antony J M. 1995 *Aquaculture*, **138** 257-66
- [12] Lakshmanan P T, Varma P R and Iyer T S 1993 Food Control 4 159-64
- [13] Boonmar S, Morita Y, Pulsrikarn C, Chaichana P, Pornruagwong S, Chaunchom S, Sychanh T, Khounsy T, Sisavath D, Yamamoto S et al 2013 *J. Global Antimicrob. Resist* **1** 157-61
- [14] Nguyen D T, Kanki M, Do Nguyen P, Le H T, Ngo P T, Tran D N, Le NH, Van Dang C, Kawai T, Kawahara R et al. 2016 *Int. J. Food Microbiol.* **236** 115-122

Acknowledgments

This study was funded by the Junior Research of Faculty of Fisheries and Marine Grant, Universitas Airlangga, 2016.