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# Coleus cultivars (Solenostemon scutellarioides (L.) Codd.) as potential bioindicators of chronic ozone exposure 

M Padri and C Umponstira<br>Natural Resources and Environment Department, Naresuan University, Phitsanulok, 35000, Thailand<br>Email: biologiunm10@gmail.com


#### Abstract

Sensitivity of plant under ozone exposure can indicate its potency of being important bioindicator. Early studies have found that coleus plant (Solenostemon scutellarioides (L.) Codd.) has a potency to be ozone bio-indicator. This study aims to investigate the effect of chronic ozone exposure on root and leaf biomass and to quantify any change based on the plant appearance. Four different cultivars of coleus plant with different colour namely fully green (FG), green purple (GP), yellow purple (YP), and reddish (RD) had been selected. These four cultivars were fumigated with three different concentrations of ozone gas ( $\leq 10 \mathrm{ppb}, 40 \mathrm{ppb}$, and 150 ppb ) for 8 hours fumigation during 30-day period of experiment. All cultivars showed a stable leaf biomass after 30-day period of ozone exposure. Similarly, root biomass of all cultivars was not significant changed after fumigation period. Nonetheless, magnitude of ozone symptoms on leaf showed variation in certain cultivars. FG cultivar showed a significant number of chlorosis leaves under 40 and 150 ppb ozone exposure. The exposure of 150 ppb ozone also caused a noticeable percentage of curling leaves on RD cultivar. Moreover, the purple area in YP and GP showed a larger ratio under exposure of 150 ppb ozone. Consequently, coleus plants displayed resistance responds in terms of biomass. On the other hand, the plants also revealed variation in leaf symptom magnitudes and colour patterns under ozone exposure. However, these cultivars are potential to be bio-indicator due to their sensitivity in terms of appearance.


## 1.Introduction

Tropospheric ozone level tends to increase in current years. Regionally, ozone was observed to increase in many places [1]. Massive growth of human population and severe managed environment are among the causes of the increase. It is important to note that in some places ozone level can be very dangerous and over the air quality standards during daytime [2]. This pollutant is shortly being the third most important greenhouse gas occurs in the ambient air [3].

This pollutant regards as cause of many decreases in cropping plants productivity [4]. Chronic symptoms and diseases after inhaling specific amount of ozone ambient have been reported in soybean [5], mug bean [6], rice [7], and many other cropping plants [8]. Mitigation about this problem is hardly conducted to its state as a secondary pollutant in which it can be sourced from many sites far from its high-level occurrences [9] unless there is the great strategies on the cropping plants to reduce its impacts [10]. The ozone exposure also affects human health [3, 11]. It can affects skin lipid conditions [12], increasing the prevalence of premature death [13] and other harmful effects [14]. Peñuelas, Ribas
[15] assumed that bioindicator can be the indication of the increase to draw comprehensive picture of this airborne problem.

Among the specific purpose of bio indicator is to use the biota to be indicator for change and/or specific case of detrimental condition in the environment or certain occasions [16]. Several plants have been proven good bio-indicators while others only showed clear sign of ozone increase after acute exposure [17-19]. The bioindicator of ozone should be sensitive of the ozone exposure and can show clear specific symptoms of high concentration of ozone exposure [20]. Moreover, the bioindicator plants are also expected to increase people awareness about the climate change and health impacts regarding this pollutant [21]

Previous researches had been conducted to assess potential plant for bio indicator purposes. Cutleaf coneflower (Rudbeckia laciniata) plants was assessed to obtain more comprehensive result of the plant regarding its visible foliar injury occurrences in which it was found to be relatively slow to occur [22] although the plant showed stippling chlorosis and necrosis and recognized as a bioindicator [23]. Ipomoea nil (L.) Roth. cv. 'Scarlett O’Hara' was also evaluated and it showed the noticeable visible leaf injuries while haziness in carbon assimilation and stomatal conductance parameters under the exposure of above 40 ppb ozone (AOT40) as the threshold [24].

However, it is rare to found some species with different cultivars can have such a major difference in terms of responses. Among those species Phaseolus vulgaris showed different responses within cultivars because of its different physiological process in their metabolisms [25].

Coleus has been recognized as the ornamental plant for many years. This plant has wide range of colors in its leaf. It is easy to find and cultivate so that this species is among the favorite ornamental plants in tropical and subtropical areas. Because of its fast growing in cultivation and captivating colors, this plant has been assessed under many stresses to obtain the most suitable cultivars in different conditions. Several studies conducted showed that this plant has fast and obvious responses to the environment changes although those responses do not lead to the death of the plant. Based on these sensitivities, several studies applied the stress for many purposes such as bioactive compound production from this plant [26] and quest for plant model for better understanding of this plant responses under environmental stresses [27-29].

Although many studies had been done to find suitable and susceptible plant for bioindicator, lac of investigation has been done on ornamental plant. Coleus has been chosen for this study as a plant model for ozone bioindicator. Thus, the aims of this study were effect of chronic ozone exposure on root and leaf biomass and to quantify any change based on the plant appearance. However, the sensitivity level and susceptibility of several cultivars of coleus was investigated and the plant can be beneficial for indicating ozone level and tis severity. Moreover, result of this study can initiate related studies especially on physiological and molecular mechanism of plant under ozone exposure.

## 2.Materials and Methods

### 2.1. Plant preparation and ozone exposure

Four cultivars of Coleus (Solenostemon scutellarioides (L.) Codd) which are Coleus Kong Green (fully green/FG), Coleus Kong Scarlet (green and purple/GP), Coleus Wizard Pastel (yellow and purple/YP), and Coleus Wizard Scarlet (reddish/RD) were chosen to be the experimental plants in this study (Figure 1). Plants were grown and treated as described in [30].

### 2.2. Ozone exposure

Chronic episode of ozone exposure was fumed into chambers of ozone fumigation as shown in Figure
2. Briefly the mechanism of the chambers is described. An ozone generator )Oz model 3020 from Ebase corp. ltd. ( was used to generate ozone gas into fumigation chambers. The ozone gas produced by the ozone generator went through a hose to be combined with air coming from an air compressor. The air passed through a charcoal filter. A valve near the end of the hose adjusts the amount of ozone that was combined with the filtered air in a plastic pipe leading to a fumigation chamber, to regulate
the ozone concentration entering the fumigation chamber. The valve was controlled by an Ozone Monitor Model 1008-PC from Dasibi Environmental Corp. to control the ozone concentration in the fumigation chambers.


Figure 1. Experimental plants (a) yellow purple cultivar (b) green cultivar (c) reddish cultivar (d) green purple cultivar.


Figure 2. Installation of ozone fumigation chambers and plant placement. (a) valve, (b) lamp, (c) door, (d) controller, (e) air Conditioner, (f) air compressor, (g) ozone generator, (h) ozone monitor, (i) air intake, (j) plants pot.

The fumigation chambers which were used in this experiment have the proportions 121 cm in length, 71 cm width and 122 cm in height. All six chambers was fumed to have three different ozone concentrations. Two chambers had charcoal-filtered air only with ozone concentration lower than 10 part per billion ( ppb ) ( $<10 \mathrm{ppb}$ ozone), two chambers contained air with ozone concentration about 40 $\mathrm{ppb}) 40 \mathrm{ppb}$ ozone(, and two chambers contained air with ozone concentration of about 150 ppb )150 ppb ozone(. Before fumigation occurs, each chamber was filled with air at the required ozone concentration, which was monitored by the ozone monitor and continued to measure during the fumigation. Ozone concentration in the chamber tended to stable during the fumigation period as shown in Figure 3.

All the other factors namely temperature and light intensity were adjusted to be similar. Temperature inside the chambers was ranging in $28-32^{\circ} \mathrm{C}$ from 06.00 to 18.00 and $25-28^{\circ} \mathrm{C} 18.00$ to 06.00 . Photosynthesis photon flux density )PPFD (inside the chamber was 400 to $500 \mu \mathrm{~mol} \mathrm{~s}^{-1} \mathrm{~m}^{-2}$ at the shortest coleus plants height for 12 hours per day. Fumigation period was 28 days with chronic ozone regime in the chamber atmosphere for 8 hours from 08.00 to 6.00 .

### 2.3. Plant evaluations and data analysis

2.3.1. Specific leaf area. Specific leaf area was measured by following the method from Wilson, Thompson [31]. Leaf area the upper fully expanded leaves of each plant was measured using Portable Leaf Area Meter model AM350 from Opti-Science. After measuring the leaf area, the leaf was dried in oven at $70^{\circ} \mathrm{C}$ for 72 hours. Dry weight was measured after drying process and the Specific Leaf Area (SLA) was calculated by dividing total area of leaf $\left(\mathrm{mm}^{2}\right)$ by total weight of the leaf (gr).
2.3.2. Root biomass. Root biomass was calculated by measuring the dry root mass. All the plants were gently pulled out with all the soil from the pots. The soil was gradually removed from the root in the running tap water. After all the materials were run out from the root the root then dried using tissue. The roots were then put into the envelope and sealed with label. Roots were dried at $70^{\circ} \mathrm{C}$ for 72 hours. The roots were taken one by one from the oven to get rid of gaining any water absorption. After that the root were measured in weight in order.
2.3.3. Symptom and purple area quantification. Every symptom was assessed quantitatively. Previous study was reported that curling leaf in RD cultivar and chlorosis in FG cultivar [30]. In this analysis, the result was an individual percentage of visible symptoms obtained from Equation (1).

$$
\begin{equation*}
\mathrm{p}=\mathrm{n}\left(\mathrm{~N}^{-1}\right) \cdot 100 \% \tag{1}
\end{equation*}
$$

$p, n$, and $N$ are percentage of symptoms, number of fully expanded damage leaves, and total number of fully expanded leaves, respectively. Percentage of number damage in each plant in the three different ozone regime thus collectively compared.

Table 1. Classification of purple area in percentage and unit of purple area.

| Broad level | Percentage | Unit of Purple <br> Area |
| :--- | :---: | :---: |
| Little area | $) 0-5 \%($ | 1 |
| Small area | $5-20 \%($ | 2 |
| Medium area | $) 21-40 \%($ | 3 |
| Medium Large area | $) 41-60 \%($ | 4 |
| Large area | $) 61-80 \%($ | 5 |
| Extra Large area | $) 81-100 \%($ | 6 |

As purple area of GP and YP cultivars (Figure 4) dramatically expanded, a quantitative investigation was conducted to draw the effect of the exposure in quantitative way using the unit proposed by Ladd, Skelly, Pippin, \& Fishman (2011). Every fully expanded leaf was classified into Purple area Unit based on the percentage of the purple area appeared in the leaves (Table 1). The data from each cultivar under three different ozone concentration were analyzed to draw the effect.

## 3.Result and discussion

### 3.1. Ozone concentration

The production of ozone gas had been monitored through 30-day period of fumigation. Fluctuation of Gas concentration was detected in every concentration (Figure 3). In the $<10 \mathrm{ppb}$ ozone chambers, the fluctuation was detected with minimum concentration 2 and maximum 15 ppb . Similarly, high range of fluctuation was found in 40 ppb ozone chambers, where the lowest was 15 ppb and the highest concentration was 58 ppb . In the highest fumigation concentration, 150 ppb ozone, the lowest concentration in this third concentration was 134 ppb whereas the ozone level was once monitored up to 173 ppb in a day. However means of all chambers, $<10 \mathrm{ppb}, 40 \mathrm{ppb}$, and 150 ppb were relatively close to the desired concentration, $7.7 \mathrm{ppb}, 40.4 \mathrm{ppb}$, and 150.8 ppb , respectively.


Figure 3. Ozone concentration inside the chamber during 30-day fumigation using MA 0.3.

### 3.2. Biomass

3.2.1. Root biomass. The assessment showed there was no significant change among the treatments )Figure 5(. It was also found that YP and RD are lighter than GP and GP. In line with that, the strucutre of the leaves also showed a similar pattern where the former cultivars showed ticker leaves whereas the latter two had thinner leaves (Figure 6). Overall, the ozone did not affect the root weight in all cultivars after 30-day period of ozone exposure. Several previous studies showed similar result. In Calluna vulgaris, there was no response in term of root biomass or length toward ozone exposure [32]. However, cropping plant such as rice responded the exposure differently as Ariyaphanphitak, Chidthaisong [33] revealed that root biomass of rice decreased when ozone exposure increased.

As ozone damage the plants in photosynthesis pigments, the reactive oxygen species derived from ozone are possible to interfere the transportation of photosynthate to belowground biomass [34].

However, this result is still premature to be widely accepted in many related species. Pregitzer, Burton [35] had emphasized that chronic exposure on perennial plants, the effect is not significant on plants root biomass. The mechanism was further discussed by Grantz, Gunn [36] where an increase of ozone is potential to induce a loss in plants' root mostly but since there is a separation effect occasionally of ozone to alter the allocation of substance to root without suppressing the productivity or photosynthesis substrate availability. Moreover, they also explained that there will be always specific cases where the effect can occurs without suppressing the translocation of photosynthesis product distribution.


Figure 4. Root Biomass of four coleus cultivars after 30-day period of fumigation.
3.2.2. Specific leaf area. This parameter aimed to describe the biomass of the leaf. Thus, it has been assessed in order to detect the change in the leaf weight per area as a response to ozone exposure. There were two cultivars with thin leaves while the other two were thicker. Thick leaf cultivars consisted of FG and GP though thin leaf cultivars consisted of YP and RD. In other parameters, these two groups did not make any significant different in terms of responses except FG which showed an increase of specific leaf area after the exposure.


Figure 5. Specific leaf area of four coleus cultivars after 30-day period of fumigation. ns: there is no significant difference among treatments.

Thickness of leaves in specific leaf area in GP, YP, and FG seems to be similar with result of Calluna vulgaris that shows a stable net photosynthesis in leaves after fumigation after ozone exposure [32] while several thin-layer leaves grasses showed similar results with FG cultivar [37]. However, it is rare to observe same species can show such noticeable different among its cultivars. The different response in FG can be described as an alternative response to the ozone since it does not contain significant amount of anthocyanin in its leaves [30] since anthocyanin acts as antioxidant to reduce detrimental effects of ozone intake [38].

### 3.3. Plant symptoms, appearances and their quantification

3.3.1. Purple area percentage in purple-area-containing cultivar. Prior study of Padri and Umponstira [30] discovered an increase of the purple area in terms of percentage per total leaf area. In this study, the purple area were quantitatively measured and further compared. It has been found that a significant increase of purple area occurred in 150 ppb ozone (Figure 6) in the cultivar where the purple area could be observed (GP and YP).


Figure 6. YG (upper) and PG (lower) cultivars after 30-day fumigation of ozone left side is the $<10 \mathrm{ppb}$ ozone and right side 150 ppb ozone. P indicates the purple area.

Wider purple areas in two cultivars YP and GP were suspected to appear because of an increase of several pigments contained in the leaves as it was prior described by Padri and Umponstira [30]. Even though percentage purple area is strongly related to the complicated genetic materials which can be induced by several stress such as light [39], anther, gibberellin, sugar content [40] and gene availability [41], ozone currently becomes among the considered causes of the change [42].

A wider purple area in leaves is caused by coloration of anthocyanin, carotenoid, and betalains [43, 44]. Moreover, it has been emphasized that there is a relation between the colours of leaves especially in the red-purple area as well as its width with the anthocyanin content [44]. Thus, the width of such colours is the units that determined by the pigments composition. As it is well-known that the pigments affects directly the colour of leaves [45], the ozone exposure in this result was the sole source that can cause the expansion of green area. In these two specific cultivars, it has been observed that there were a significant increase of purple area in the 150 ppb ozone exposure. Thus, this result can comply the prior research regarding these two cultivars in which the contents of purple pigments was increasing after ozone exposure [30].


Figure 7. Ratio unit of purple area on the leaf of four coleus cultivars after 30-days fumigation.
3.3.2. Curling and chlorotic area. Statistically, there was no different between $<10 \mathrm{ppb}$ ozone and 40 ppb ozone on curling leaf symptom in RD where the curling leaf symptom was observed. Nevertheless, 150 ppb ozone was showing a significant different result from other treatments (Figure 8). Nevertheless, FG showed a different response in 40 ppb ozone and 150 ppb ozone compared with CF (Figure 8). It is necessary to note that curling leaves showed an increase in each additional ozone concentration. The percentage of chlorotic leaves was starting to increase at 40 ppb ozone although the statistical analysis does not show any significant difference. Nonetheless, the change had large amount of chlorotic leaves in 150 ppb ozone The chlorotic and curling leaf are common symptoms of leaf under ozone exposure caused by the rupture of the plastid containing chlorophyll and excess production of ethylene under oxidative stresses, respectively [46-48].


Figure 8. Quantification of symptoms on leaf of FG and RD coleus cultivars after 30-days period of ozone exposure.

As the most famous method to assess the air pollution effect on plants, percentage of leaf area that affected have been estimated in many plants [49] even though to some extent it can be no reliable due to the high probability of bias [50]. Even so, a clear pattern of different percentage of both symptoms can rise the reliability of this estimation method. the only warning toward this method is when it comes to the lage survey and the bias can occur from many surveyors [51]. Lastly, this method which called Horsfall-Barratt rating system from Barratt and Horsfall [52] is still realible in the laboratory and specific field assessment for ozone symptoms.

### 3.4. Bioindicator potential

Bioindicators are organisms or communities in which the response of certain change and/or occurrence of novel process observed to comprehensively draw the condition of the ecosystem and or environment [53]. In ozone, it can range from forbs to trees [54]. Nevertheless, the term of bioindicator nowadays is not limited only to draw the ecosystem condition but also to specifically point the change of environmental state at defined time [55]. In the nature, plants respond elevation ozone in many different physiological and thus affecting the morphological changes such as stipples, patches, discolored, and many more in which some of them are specific and unspecific symptoms [17]. However, apart from the curling and chlorotic leaf symptoms, the change of morphological appearance specifically the purple area shows state of the art in symptoms of the leaf where most of the passive bioindicator of ozone show leaf injuries as the main symptoms [17, 22, 23, 55].
Since the novel and valid change of the plants in terms of ozone responses occurred, the degree of responses at efficient level of ozone exposure is also an important part of bioindicator plant [20]. According to Sandermann Jr [56] who proposed the level of sensitivity, all of the cultivars showed in intermediate sensitivity to ozone exposure. Yet, for bioindicator purpose, FG and RD are the potential cultivars. For bio indicator purposes, Manning, Godzik [57] stated that bio-indicator of ozone can be chosen by considering the visible symptoms that easily recognized and thus this requirement was also demostrated in the results.

Several plant species have been showing a sensitivity to ozone exposure and later on it is called ozone-sensitive plants [58, 59]. The plants sometimes make a big different and show several symptoms and loose of biomass after or during ozone exposure. On the other hand, even in the most sensitive plants, the effects of ozone sometimes are very nebulous and the symptoms end up being neglected due to its low appearance. However, performance of this plants shows a vast response under moderate exposure of ozone and thus it can be categorize as potential bioindicator plant [54]. Further, physiochemical activities of this plant under such stress condition will be able to reveal more in terms of metabolic pathway that affected.

## 4.Conclusion

Biomass parameters namely root biomass and specific leaf area showed a stable numbers after the assessment whereas the scale of ozone symptoms and its occurrence on leaf showed different quantity in specific cultivars. FG cultivar displayed chlorosis leaves under 40 and 150 ppb ozone exposure while RD presented the same significant change in terms of curling leaf under 150 pb ozone exposure. Purple area of GP and YP were increasing as the ozone did at 150 ppb ozone level. To conclude, FG and YP cultivars are potential plants for bio-indicator based on their performance in response to the ozone exposure. Correspondingly, as this result reveals a very promising respond to ozone stress, further study is necessary to confirm this result in terms of molecular effect on this plant.

## 5.References

[1] Leventidou E, Weber M, Eichmann K U, Burrows J P, Heue K P, Thompson A M, et al. 2018 Harmonisation and trends of 20-year tropical tropospheric ozone data Atmospheric Chemistry \& Physics 18 9189-205
[2] Cooper O R, Parrish D, Ziemke J, Cupeiro M, Galbally I, Gilge S, et al. 2014 Global distribution and trends of tropospheric ozone: An observation-based review Elem Sci Anth 29 1-28
[3] Guicherit R, Roemer M 2000 Tropospheric ozone trends Chemosphere - Global Change Science 2 167-83
[4] McGrath J M, Betzelberger A M, Wang S, Shook E, Zhu X G, Long S P, et al. 2015 An analysis of ozone damage to historical maize and soybean yields in the United States Proceedings of the National Academy of Sciences 112 14390-5
[5] Sun J, Feng Z, Ort D R 2014 Impacts of rising tropospheric ozone on photosynthesis and metabolite levels on field grown soybean Plant Science 226 147-61
[6] Chaudhary N, Agrawal S 2015 The role of elevated ozone on growth, yield and seed quality amongst six cultivars of mung bean Ecotoxicology and environmental safety 111 286-94
[7] Tsukahara K, Sawada H, Kohno Y, Matsuura T, Mori I C, Terao T, et al. 2015 Ozone-induced rice grain yield loss is triggered via a change in panicle morphology that is controlled by Aberrant Panicle Oorganization 1 gene PLoS One 10 1-14
[8] Ingvordsen C H, Backes G, Lyngkjær M F, Peltonen-Sainio P, Jensen J D, Jalli M, et al. 2015 Significant decrease in yield under future climate conditions: Stability and production of 138 spring barley accessions European Journal of Agronomy 63 105-13
[9] Tong L, Zhang H, Yu J, He M, Xu N, Zhang J, et al. 2017 Characteristics of surface ozone and nitrogen oxides at urban, suburban and rural sites in Ningbo, China Atmospheric Research 187 57-68
[10] Calfapietra C, Peñuelas J, Niinemets Ü 2015 Urban plant physiology: adaptation-mitigation strategies under permanent stress Trends in plant science 20 72-5
[11] Karlsson P E, Klingberg J, Engardt M, Andersson C, Langner J, Karlsson G P, et al. 2017 Past, present and future concentrations of ground-level ozone and potential impacts on ecosystems and human health in northern Europe Science of The Total Environment 576 2235
[12] Lakey P S, Wisthaler A, Berkemeier T, Mikoviny T, Pöschl U, Shiraiwa M 2017 Chemical kinetics of multiphase reactions between ozone and human skin lipids: Implications for indoor air quality and health effects Indoor air 27 816-28
[13] Shindell D, Kuylenstierna J C, Vignati E, van Dingenen R, Amann M, Klimont Z, et al. 2012 Simultaneously mitigating near-term climate change and improving human health and food security Science 335 183-9
[14] Fann N, Nolte C G, Dolwick P, Spero T L, Brown A C, Phillips S, et al. 2015 The geographic distribution and economic value of climate change-related ozone health impacts in the United States in 2030 Journal of the Air \& Waste Management Association 65 570-80
[15] Peñuelas J, Ribas A, Gimeno B, Filella I 1999 Dependence of ozone biomonitoring on meteorological conditions of different sites in Catalonia (NE Spain) Environmental Monitoring and Assessment 56 221-4
[16] Schreck E, Sarret G, Oliva P, Calas A, Sobanska S, Guédron S, et al. 2016 Is Tillandsia capillaris an efficient bioindicator of atmospheric metal and metalloid deposition? Insights from five months of monitoring in an urban mining area Ecological indicators 67 227-37
[17] Moura B B, de Souza S R, Alves E S 2014 Response of Brazilian native trees to acute ozone dose Environmental Science and Pollution Research 21 4220-7
[18] Akkoyunlu E, Tıpırdamaz R, Başaran S, Sarıbaşak H, Özkum D, Güllü G. 2015 Ambient Ozone Levels in the Eastern Mediterranean Region and Assessment of Its Effect on the Forested Mountain Areas of Southern Turkey. Plants, Pollutants and Remediation (Dordrecht: Springer) 2015. p. 349-60.
[19] Agathokleous E, Saitanis J, Satoh F, Koike T 2015 Wild plant species as subjects in $\mathrm{O}_{3}$ research Eurasian Journal of Forest Research 18 1-36
[20] Edwards C, Subler S, Chen S, Bogomolov D, Straalen N, Krivolutsky D 1996 Essential criteria for selecting bioindicator species, processes, or systems to assess the environmental impact of chemicals on soil ecosystems Bioindicator systems for soil pollution 67-84
[21] Markert B A, Breure A M, Zechmeister H G 2003 Bioindicators and Biomonitor (Netherland: Elsevier)
[22] Pringle J, Yu C, Sachs M, Ellis R 2018 Assessing ozone damage to cutleaf coneflower in an ozone bioindicator garden Journal of the Franklin Institute 355 6152-68
[23] Neufeld H S, Johnson J, Kohut R 2018 Comparative ozone responses of cutleaf coneflowers (Rudbeckia laciniata var. digitata, var. ampla) from Rocky Mountain and Great Smoky Mountains National Parks, USA Science of the Total Environment 610 591-601
[24] Moraes R M, Meirelles S T, Silva S F, Silva D T, de Assis P, Viola S 2014 Leaf injury and gas exchange in Ipomoea nil cv. Scarlett O'Hara, an ozone bioindicator species, in Sao Paulo, Brazil Atmospheric Pollution Research 5-12
[25] Villányi V, Ürmös Z, Turk B, Batič F, Csintalan Z 2014 Photosynthesis of ozone-sensitive andresistant Phaseolus vulgaris genotypes under ambient ozone and moderate heat stress Photosynthetica 52 604-13
[26] Nusrath A, Ramadas D 2018 Quantitative analysis of chemical constituents in medicinal plant coleus aromaticus extracts International Journal of Research in Medical Sciences 6 10021005
[27] Hu M, Yuan J 2018 Responses of Chemical Functional Groups in Coleus Blumei Roots under Lead Stress with Exogenous Selenium Treatment Using 2-dimensional Correlation FTIR Spectra Polish Journal of Environmental Studies 27 2561-72
[28] Kotagiri D, Kolluru VC 2017 Effect of Salinity Stress on the Morphology and Physiology of Five Different Coleus Species Biomedical and Pharmacology Journal 10 1639-49.
[29] Ibrahim K M, Musbah H M 2018 Increasing poly phenols in Coleus blumei at the cellular and intact plant levels using PEG stress Research Journal of Pharmacy and Technology 11 3217
[30] Padri M, Umponstira C 2017 Physiological and Morphological Responses to Ozone Exposure of Coleus (Solenostemon scutellarioides (L.) Codd) International Journal on Advanced Science, Engineering and Information Technology 7 2131-8
[31] Wilson P J, Thompson K E N, Hodgson J G 1999 Specific leaf area and leaf dry matter content as alternative predictors of plant strategies New Phytologist 143 155-62
[32] Foot J, Caporn S, Lee J, Ashenden T 1996 The effect of long-term ozone fumigation on the growth, physiology and frost sensitivity of Calluna vulgaris New Phytologist 133 503-11
[33] Ariyaphanphitak W, Chidthaisong A, Sarobol E, Bashkin V, Towprayoon S 2005 Effects of elevated ozone concentrations on Thai jasmine rice cultivars (Oryza sativa L.) Water, Air, and Soil Pollution 167 179-200
[34] Olszyk D M, Wise C 1997 Interactive effects of elevated CO 2 and O 3 on rice and flacca tomato Agriculture, ecosystems \& environment 66 1-10
[35] Pregitzer K S, Burton A J, King J S, Zak D R 2008 Soil respiration, root biomass, and root turnover following long-term exposure of northern forests to elevated atmospheric CO 2 and tropospheric O3 New Phytologist 180 153-61
[36] Grantz D A, Gunn S, Vu H B 2006 O3 impacts on plant development: a meta-analysis of root/shoot allocation and growth Plant, Cell \& Environment 29 1193-209
[37] Franzaring J, Tonneijck A E G, Kooijman A W N, Dueck T A 2000 Growth responses to ozone in plant species from wetlands Environmental and Experimental Botany 44 39-48
[38] Treutter D 2006 Significance of flavonoids in plant resistance: a review Environmental Chemistry Letters 4147
[39] Nakatsuka A, Yamagishi M, Nakano M, Tasaki K, Kobayashi N 2009 Light-induced expression of basic helix-loop-helix genes involved in anthocyanin biosynthesis in flowers and leaves of Asiatic hybrid lily Scientia Horticulturae 121 84-91
[40] Weiss D 2000 Regulation of flower pigmentation and growth: Multiple signaling pathways control anthocyanin synthesis in expanding petals Physiologia Plantarum 110 152-7.
[41] Boye C L 1941 An allelic series inColeus Journal of Genetics 42191
[42] Carter G A, Knapp A K 2001 Leaf optical properties in higher plants: linking spectral characteristics to stress and chlorophyll concentration American journal of botany 88 677-84
[43] Lebowitz RJ 1985 The genetics and breeding of coleus Plant Breeding Reviews, Volume 3 34360
[44] Nguyen P, Cin V D 2009 The role of light on foliage colour development in coleus (Solenostemon scutellarioides (L.) Codd) Plant Physiology and Biochemistry 47 934-45
[45] Vollenweider P, Günthardt-Goerg M S 2005 Diagnosis of abiotic and biotic stress factors using the visible symptoms in foliage Environmental Pollution 137 455-65
[46] Kubo A, Saji H, Tanaka K, Kondo N 1995 Expression of Arabidopsis cytosolic ascorbate peroxidase gene in response to ozone or sulfur dioxide Plant molecular biology 29 479-89
[47] Sharma Y K, Davis K R 1994 Ozone-induced expression of stress-related genes in Arabidopsis thaliana Plant Physiology 105 1089-96
[48] Gravano E, Bussotti F, Strasser R J, Schaub M, Novak K, Skelly J, et al. 2004 Ozone symptoms in leaves of woody plants in open-top chambers: ultrastructural and physiological characteristics Physiologia Plantarum 121 620-33
[49] Chester K S 1959 How sick is the plant Plant pathology 1 99-142
[50] Knudson L L, Tibbitts T W, Edwards G E 1977 Measurement of ozone injury by determination of leaf chlorophyll concentration Plant physiology 60 606-8
[51] Bussotti F, Schaub M, Cozzi A, Kräuchi N, Ferretti M, Novak K, et al. 2003 Assessment of ozone visible symptoms in the field: perspectives of quality control Environmental Pollution 125 81-9
[52] Barratt R, Horsfall J 1945 An improved grading system for measuring plant disease Phytopathology 35655
[53] Gerhardt A 2002 Bioindicator species and their use in biomonitoring Environmental monitoring 177-123
[54] Manning W J, Godzik B, Musselman R 2002 Potential bioindicator plant species for ambient ozone in forested mountain areas of central Europe Environmental Pollution 119 283-90
[55] Nakazato RK, Esposito MP, Cardoso-Gustavson P, Bulbovas P, Pedroso ANV, de Assis PILS, et al. 2018 Efficiency of biomonitoring methods applying tropical bioindicator plants for assessing the phytoxicity of the air pollutants in SE, Brazil Environmental Science and Pollution Research 25 19323-37
[56] Sandermann Jr H 1996 Ozone and plant health Annual review of phytopathology 34 347-66
[57] Manning W, Godzik B, Musselman R 2002 Potential bioindicator plant species for ambient ozone in forested mountain areas of central Europe Environmental Pollution 119 283-90
[58] Burkey K O, Booker F L, Ainsworth EA, Nelson RL 2012 Field assessment of a snap bean ozone bioindicator system under elevated ozone and carbon dioxide in a free air system Environmental pollution 166 167-71
[59] Bergweiler C, Carreras H, Wannaz E, Rodriguez J, Toselli B, Olcese L, et al. 2008 Field surveys for potential ozone bioindicator plant species in Argentina Environmental monitoring and assessment 138 305-12

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